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Some alkaloids and flavonoids from *Cissampelos capensis*

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Following the screening of several plant species from an inventory of common medicinal plants from South Africa for medicinal properties, *Cissampelos capensis* was selected for further investigation due to its interesting and useful ethnomedicinal properties. This study attempts to relate specific constituents present in this plant with its widespread ethnomedicinal uses. Six compounds were isolated and their structures were unambiguously established by spectroscopic methods. The compounds are: 5,6-dehydro-4,5-dihydroxy-1,3,6-trimethoxy-17-methylmorphinan-7-one (1); 1,2-methylenedioxy-3-hydroxy-9,10-dimethoxyaporphine (2); 5,6-didehydro-4-hydroxy-3,6-dimethoxy-17-methylmorphinan-7-one (3); 3,7,8,3'-tetramethoxy-6-C-methyl-5,4'-dihydroxyflavone (6-C-methylquercetin 3, 3',7, 8-tetramethyl ether) (4); 5, 7, 8-trihydroxy-2', 5'-dimethoxy-3',4'-methylenedioxyisoflavanone (5); 3-methoxy-6-C-methyl-3',4',5,7,8-pentahydroxyflavone (6-C-methylquercetin-3-methyl ether) (6). Five of the isolated compounds, (viz., 1,2,4,5 and 6) have, to our knowledge, not been reported previously. The crude fractions and isolates were tested for cytotoxicity using the brine shrimp lethality test and for antimicrobial properties using nine microbes, including three Gram -ve, three Gram +ve bacteria and three fungi. The Gram-negative bacteria were *Pseudomonas aeruginosa* (NCTC 10332), *Proteus vulgaris* (NCTC 4175) and *Escherichia coli* Sero type 1 (NCTC 09001), while the Gram-positive bacteria were *Bacillus subtilis* (NCTC 8236), *Staphylococcus aureus* (NCTC 13134) and *Bacillus licheniformis* (NCTC 01097). The Fungal species used were *Candida albicans* (ATCC 90028), *Candida eropiralis* (ATCC 750) and *Aspergillus niger* (ATCC 10578). The n-Hex fractions were not active while the highest activities were found in the methanolic extracts. The total tertiary alkaloid fraction (TTA) showed the highest activity against the *Bacillus subtilis*. Compounds 1, 2 and 5 appear to be the most promising with regards to the prospects of drug development.

Key words: Phytochemistry, *Cissampelos capensis*, antimicrobial, cytotoxicity, isoflavanone, aporphine, morphinandienone.

INTRODUCTION

Medicinal plants, in a variety of forms have been recorded to cure, manage and control conditions in people for at least four thousand years, and it may be

assumed that this practice extends even longer (Christophersen et al., 1991). The World Health Organisation (WHO) has recognized this and has accepted

it as an essential building block for primary health care (Akerle, 1988; WHO, 2005). The vibrant healing power of *C. capensis* has been recognized in recent times (Van and Gericke, 2000; Iwu, 1993; WHO, 2002), and especially by traditional healers in South Africa, who are known by the Zulu people as inyangas, herbalists, isangomas or diviners while they are also known as amaxhwele, amagqira (Xhosa), ngaka (Sotho), nanga, mungome or maine (Vhavenda) (Van, et al., 1997; Mander, 1998; De Wet and Van, 2008; Van, 2008; Van et al., 2008).

The plant is one out of 520 species in 75 genera (Van et al., 1997), which belongs to the family of menispermaceae. It is widely distributed in the sandy slopes and scrublands of the Northern, Western and Eastern Cape Provinces of South Africa and northwards into Namibia. It is called by different local name such as Dawidjiewortel (Afrikaans) and Mayisake (Xhosa). Due to its richness in bisbenzylisoquinoline alkaloids, this family is used worldwide in traditional medicine to treat a variety of ailments especially cancer (Barbosa et al., 2000; DeWet et al., 2004, 2005; DeWet, 2006; Watt and Breyer-Brandwijk, 1962; Smith, 1966; Rood, 1994; Cillie, 1992; Dykman, 1891; Iwu, 1993). It is a woody dioecious perennial climbing vine with a twining stem, shrublet without tendrils and supports itself by twining around the stems of other plants, fence or walls. The leaves are rounded, bright green, almost without hairs. The flowers, which usually sprout between February and May, are axillary with velvety-hairy and greenish in colour. The fruit is inedible, dark, grape-sized berries. The root may be up to 2.5cm in diameter with a grey-brown bark (Botha, 1980). The plant is usually confused with *Zehneria scabra* (Cucurbitaceae), which in some parts is known by the same local name (Smith, 1966).

The latter resembles a cucumber and can be distinguished by the spirally coiled tendrils. Nearly all parts of the plant are used; from the whole vine, seed, bark, leaf to the root. It is traditionally used in South Africa to treat a variety of ailments such as dysentery, menstrual problems, prevention of miscarriage, cholera, colic, snakebite, measles, fever, diabetes, tuberculosis, stomach and skin cancers, etc (Watt and Breyer-Brandwijk, 1962; Rood, 1994; Van Wyk and Gericke, 2000; VanWyk et al., 1997; De Wet and Van Wyk, 2008; Barbosa- Filho et al., 2000; De Wet et al., 2004, 2005). In 1962, it was reported that *C. pareira* demonstrated anti-inflammatory, smooth muscle relaxant, antispasmodic, diuretic effect and uterine relaxant actions in various laboratory animals and this was further documented by Pillay et al. (2008) and Taylor (2005). Ssegawa and Kasenene, (2007) described its antiulcerous actions and mild hypoglycemic activity. Further studies have also shown that the roots described

its antiulcerous actions and shown that the roots of cissampelos species have anticonvulsant activity, antioxidant properties, antimicrobial activity, antihypertensive, antimalarial effects, antispasmodic and antitumour properties (Dic. Nat. prod., 1996,2006; Bruneton, 1995; Wu, 2007; Amresh et al., 2007; Amresh et al., 2004; Gessler et al., 1995; Ramirez et al., 2003; Sanchez et al., 2001; Taylor, 2005; Graham et al., 2000; Hamill et al., 2003). The antinociceptive and antiarthritic activity of *Cissampelos pareira* roots were also highlighted by Amresh et al., (2007) while the antifertility activity and anti-inflammatory activity were described by Ganguly et al. (2007) and Amresh et al. (2007) respectively. The Khoi-San and the Cape Dutch recognised the importance of this plant in Cape herbal medicine as one of the major distinct plants used for the treatment of most diseases (Van Wyk, 2008), while during the evaluation of some South African medicinal plants for antimalarial properties, it was discovered that *C. capensis* possesses antiplasmodial activity (Pillay et al., 2008). The medicinal values of *Cissampelos* species in the Sango bay area in Southern Uganda was also recorded by Ssegawa and Kasenene (2007).

Phytochemical screening showed the presence of alkaloids, saponins, flavonoids, phenolics and tannins while cardiac glycosides and anthraquinones were not detected in the different extracts (Babajide et al, 2010). A large number of biologically active alkaloids have been isolated from several *Cissampelos* species among these were cissampareine and magnoflorine, (Jittra et al., 2005; Freitas et al., 1995). Other alkaloids such as cissaglaberrimine and oxobuxifoline were isolated from *Cyathea glaberrima* (Barbosa-filho et al., 1997) while Milonine, an 8, 14-dihydromorphinandienone alkaloid was also isolated from the dried leaves of *Cissampelos sympodialis* (Freitas et al., 1995). Similarly, Sloan et al. (2007) showed the isolation of two known aporphine alkaloids, (S)-dicentrine and (S)-neolitsine from methanolic extract of *C. capensis*, which were found to be potent anthelmintics. To the best of our knowledge no flavonoidal compounds isolated from *C. capensis* has been published.

The present study involves a more intense investigation of the constituents present in *C. capensis* as well as their biological activities.

MATERIALS AND METHODS

General experimental procedures All laboratory grade solvents were distilled prior to use. Spectroscopic grade solvents were used as such. Adsorption column chromatography was performed using Merck Silica gel 60 H (0.040 -0.063 mm particle size, Merck).

Unless otherwise specified, a column with internal diameter of 25

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mm was used and volumes of 10 ml were collected in Pyrex test tubes. Size-exclusion column chromatography was performed using Sephadex LH-20 (Sigma) pre-swollen in the specified solvent before loading of sample.

Preparative Thin Layer Chromatography (TLC) was performed using Merck Silica gel 60 PF₂₅₄ on glass plates (20 cm x 20 cm) and with a thickness of 0.5 mm. Analytical TLC was conducted on normal-phase Merck Silica gel 60 PF₂₅₄ on pre-coated aluminium plates. Separated compounds on TLC and Preparative Thin Layer Chromatography (PTLC) plates were visualized under Ultraviolet (UV) light at (254 and 366 nm), and spraying of the plates where required were carried out using 2% vanillin in H₂SO₄ followed by heating at 110 °C for 2 to 4 mins. Quercetin, rutin and catechin were used as reference standards for flavonoids, gallic acid for tannins and oleanolic acid for essential oils. In the case of the Alkaloids, the dragendorff spraying reagent was used for visualization while quinine and berberine were used as standards.

Reversed phase TLC (RPTLC) was performed using pre-coated plates (Merck, RP-18, F254 2, 0.25 mm thickness) and spots were detected as described above. R_f values were determined at room temperature using different solvent systems as applicable. Melting points were determined on a Fisher – John's melting point apparatus (Fisher-Scientific). All melting points are uncorrected. Mass spectra were obtained using JEOL JMS-AX505HA double-focusing probe at 70 eV while UV spectra with a Unicam UV 4 -100 UV/Vis recording spectrophotometer. The spectra were recorded over the range of 200 to 550 nm, at a concentration of 0.02 mg/ml in spectroscopic grade methanol. All infra-red (IR) spectra were recorded on a Perkin Elmer universal ATR Spectrum 100 series FT-IR spectrometer while Nuclear magnetic resonance (NMR) spectra were recorded on a Bruker AMX-400 (400 MHz for ¹H and 100 MHz for ¹³C) spectrometer, using a triple resonance probe head with self-shielded gradient coils and a Bruker Z-gradient accessory delivery squared gradients. The chemical shifts are expressed in ppm relative to tetramethylsilane (TMS).

Plant

The aerial shoot and root of *C. capensis* were collected in May 2007 from the University of the Western Cape (UWC) Cape Nature Conservation Reserve, Bellville, Cape Town, South Africa, authenticated by a plant systematist and a voucher specimen [number Weitz 1056(UWC)] deposited in the herbarium of the University of the Western Cape. Preparation of the extracts was performed as described by Babajide et al. (2010).

Extraction and isolation

Each of the extracts that is, hexane (Hex), dichloromethane (DCM), ethyl acetate (EtOAc), methanol (MeOH), TTA and residual aqueous fraction (BTA) were separately evaporated under reduced pressure at 40 °C while aqueous extracts were concentrated by freeze-drying. The preliminary phytochemical screening (Wagner and Bladt, 2001) was carried out for detection of classes of secondary plant metabolites as shown in Babajide et al. (2010)

Fractionation of the TTA

The TTA (1.6g) was adsorbed on silica gel and the constituents were separated by column gradient chromatography (CGC) using an increasing gradient of Hex and CHCl₃ and finally MeOH was employed. The fractions collected were analyzed by TLC using Hex– CHCl₃ – MeOH (1: 5 : 1). Tubes showing similar TLC characteristics were bulked together and concentrated. Seven fractions coded A to G were obtained and the bioactivities of each

were monitored using the brine shrimp cytotoxicity test. Fraction E (0.30g) with LC₅₀ value of 2.4050 was selected for further fractionation. Fraction E (0.25 g) was rechromatographed using CGC to afford five fractions: TI to TV and fractions TIII (0.122g) with an LC₅₀ value of 3.0226 and TIV (0.047g) with an LC₅₀ value of 0.8655 were further investigated. Repeated preparative TLC of TIII gave compound (1) a brownish yellow powder (0.058 g) and that of TIV afforded compound (2) as a brown amorphous solid (0.021g).

Extraction and fractionation of the aerial shoot

The aerial shoot (1 kg) was sequentially extracted to afford the hexane (17.35g, 1.74%), dichloromethane (26.33g, 2.63%), ethyl acetate (32.84g, 3.28%), methanol (56.24g, 5.62%) and water extracts (25.71g, 2.57%). The methanol extract (25.6g) was chromatographed using same method as described for TTA. Ten fractions coded A to J were obtained and fraction F (1.96g) with an LC₅₀ value of 1.2332 was selected for further fractionation which afforded five fractions viz., FI to FV of which FII (0.520g) with LC₅₀ of 66.73, FIII (0.211g) with LC₅₀ of 3.442, FIV (0.115g) with LC₅₀ of 17.56 and FV (0.106g) with LC₅₀ of 45.46 were further investigated. Fraction FII (0.5 g) was chromatographed using CGC and was further purified using sephadex LH-20 and repeated preparative TLC. This gave a golden brown powder (0.04g) of compound (3). Fraction FIII (0.20 g) was also rechromatographed and further subjected to repeated preparative TLC to afford a brownish yellow powder which was recrystallized from EtOH to give a further quantity of compound (1) (a brownish yellow powder, 0.028 g) while fraction FIV (0.10 g) after preparative TLC afforded (4) as a yellow amorphous powder (0.026 g) and fraction FV (0.10 g) gave compound (5) as a yellow powder (0.024g).

Extraction and fractionation of the root

The same extraction procedure was repeated for the dried root (1 kg). This afforded a hexane (7.38g, 0.74%), dichloromethane (12.65g, 1.27%), ethyl acetate (27.19g, 2.72%), methanol (53.05g, 5.31%) and water extract (26.44g, 2.64%). The methanol extract (30.0g) was chromatographed as described for the aerial shoot. Out of the five fractions collected, fractions M (5.68g) with LC₅₀ value of 5.2375 and N (3.67g) with LC₅₀ of 19.296 were selected for further fractionation. The fractionation of M (5.0 g) by repeated CGC and monitoring of their LC₅₀ values resulted more in compounds of (1) (0.078g), (2) (0.065g) and (5) (0.088g). Similarly, fraction N (3.2 g) was rechromatographed and accorded more of compound (4) (0.068g) and a new compound viz., (6) (0.135g) a creamy yellow solid was obtained from subfractions NIII (0.73 g) with LC₅₀ of 76.112.

Dihydromorphinandienone alkaloid (1)

A reddish yellow powder with m.p. 85 to 87 °C; UV λ_{max} nm (log ε): 222, 260, 304; IR ν_{max} cm⁻¹ pronounced peaks: 3485, 2940, 1685, 1615, 1500; EIMS: molecular ion peak (M)⁺ m/z 375.0216 (calculated for C₂₀H₂₅O₆N: 375.1682). The ¹H NMR and ¹³C NMR see Table 1

9, 10 – Dimethoxyaporphine alkaloid (2)

A reddish-brown fluffy powder with m.p. 221 – 223 °C; UV λ_{max} nm(log ε): 225, 243, 277, 300 (shoulder). IR ν_{max} cm⁻¹ pronounced peaks: 3420, 3050, 2925, 1636, 1458, 1387, 1060. EIMS: molecular ion peak (M)⁺ m/z 341.0112. (calculated for C₁₉H₁₉O₅N: 341.1263). For the ¹H NMR and ¹³C NMR (Table 2).

Table 1. ^1H NMR (400 MHz) data of (1) and ^{13}C NMR (100 MHz) data of (1) in $\text{C}_5\text{D}_5\text{N}$.

C	^{13}C	^1H
1	146.4	--
2	111.2	6.91 s (1H)
3	148.4	--
4	145.6	--
5	125.3	--
6	152.2	--
7	194.7	--
8	40.3	ax 3.63 dd (J = 14.2, 17.4 Hz) eq. 2.64 dd (J = 17.4, 4.6 Hz)
9	56.9	2.83 br d (J = 5.6 Hz)
10	28.5	ax 3.12 d (J = 17.0 Hz) eq 2.74 dd (J = 17.0, 5.6 Hz)
11	131.2	--
12	128.7	--
13	38.5	--
14	42.7	2.40 m
15	32.3	2.21 – 2.11 m (2H)
16	46.5	2.40 m (1H)/2.30 m (1H)
MeO-1	57.2	3.83 s (3H)
MeO-3	56.5	3.76 s (3H)
MeO-6	55.3	3.69 s (3H)
Me-N	43.1	2.26 s (3H)
HO-4	--	10.75 br s (1H)
HO-5	--	11.50 br s (1H)

8,14-Dihydromorphinandienone alkaloid (3)

A golden brown powder with m.p 77 to 80°C. UV λ_{max} nm (log ϵ): 226, 261,302; IR ($\nu \text{ cm}^{-1}$) pronounced peaks: 3517; 2952, 1685, 1616, and 1510. EIMS: the molecular ion base peak (M^+) m/z 329.2133 (calculated for $\text{C}_{19}\text{H}_{23}\text{O}_4\text{N}$: 329.1627). The ^1H NMR and ^{13}C NMR (Table 3)

6-C-methylquercetin -3, 3',7, 8 -tetramethyl ether (4)

A yellow amorphous powder with m.p. 234 to 236°C. UV λ_{max} nm (log ϵ): 255 (4.64), 357 (4.62); +NaOAc: 271, 365; +NaOH: 276, 343, 410. + AlCl_3 : 279, 440. IR ($\nu \text{ cm}^{-1}$) pronounced peaks: 3401, 2932, 1653, 1612, 1556 EIMS: molecular ion peak (M^+) m/z 388.0103. (calculated for $\text{C}_{20}\text{H}_{20}\text{O}_8$: 388.1158). ^1H NMR: δ 2.14 (3H, s, Me-6), 3.74 (3H, s, OMe-3), 3.96 (3H, s, OMe-3'), 4.01 (3H, s, OMe-7); 4.56 (3H, s, OMe-8), 6.91 (1H, d, J= 8.5 Hz, H-5'), 7.60 (1H, dd, J = 2.3, 8.4 Hz, H-6'), 7.70 (1H, d, J= 2.3 Hz, H-2'), 10.90 (1H, s, 4'-OH), 12.92 (1H, s, 5-OH). ^{13}C NMR (Table 4)

Methylenedioxyisoflavanone (5)

A yellow waxy powder with m.p. 185 to 187°C; UV λ_{max} nm (log ϵ): 214(4.10), 218 (4.12), 288 (4.00), 333 (sh) (3.25). IR $\nu_{\text{max}} \text{ cm}^{-1}$ pronounced peaks at : 3421, 2914, 1634, 1472, 1386, 1358, 1257, 1224, 1161, 1100, 1068, 925; EIMS: molecular ion peak (M^+) m/z 378.0625 (calculated for $\text{C}_{18}\text{H}_{18}\text{O}_9$, 378.0623). ^1H NMR, ^{13}C NMR and HMBC are as shown in Tables 5 and 6 respectively.

6-C-Methylquercetin 3-methyl ether (6)

A creamy yellow powder with m.p. 217 to 219°C. UV λ_{max} nm (log

ϵ): 260 (4.62), 351 (4.60); +NaOAc: 260, 355; +NaOH: 270, 405; + AlCl_3 : 277, 435. IR ($\nu \text{ cm}^{-1}$) pronounced peaks: 3500 – 3400, 1652, 1613, 1568, 1556 EIMS: molecular ion peak (M^+) m/z 346.0752 (calculated for $\text{C}_{17}\text{H}_{14}\text{O}_8$: 346.0688). ^1H NMR: δ 2.11 (3H, s, Me-6), 3.87 (3H, s, OMe-3), 7.03 (1H, d, J=8.6Hz, H-5'), 7.62 (1H, dd, J= 2.3, 8.6 Hz, H-6'), 7.75 (1H, d, J= 2.3Hz, H-2'), 10.23 (1H, s, 4'-OH), 11.15 (1H, s, 3'-OH), 12.05 (1H, s, 8-OH), 12.90 (1H, s, 7-OH), 13.05 (1H, s, 5-OH). ^{13}C NMR: see Table 7.

Brine shrimp lethality bioassay

The brine shrimp lethality bioassay was carried out as previously described (Babajide et al., 2008, 2010) where about 2 g of the Brine shrimp eggs (*Artemia salina* Leach) were hatched in 2 L of sea water using a large plastic case as an artificially partitioned dam. The number of dead nauplii were counted and recorded (lethality data) after 24 h. The numbers of dead nauplii were used for calculating the LC_{50} at 95% confidence limit by the Finney Probit analysis program. LC_{50} values greater than 800 ppm or in that range were considered inactive.

Antimicrobial activity

The organisms used in the screening tests were as follows: The gram-negative bacteria were *P. aeruginosa* (NCTC 10332), *P. vulgaris* (NCTC 4175) and *E. coli* sero type 1 (NCTC 09001), while the gram-positive bacteria were *B. subtilis* (NCTC 8236), *S. aureus* (NCTC 13134) and *B. licheniformis* (NCTC 01097). These species are considered as the most important pathogens (NCCLS, 1990). Fungal species used were *C. albicans* (ATCC 90028), *C. eropiralis* (ATCC 750) and *A. Niger* (ATCC 10578). Cultures were grown in sabouraud dextrose (SD) broth at 37°C and maintained on SD agar at 4°C. A colony of each bacterial strain was suspended in 1 ml of

Mueller–Hinton broth and incubated for 18 h at 37°C. A subculture was again made after 6 h. The subculture was diluted 1/50 in the same broth before use. A disc diffusion assay was used to determine the inhibition of bacterial growth by the plant extracts, fractions, sub fractions and isolates (Rasoanaivo et al., 1993). Plant extracts, fractions and isolates were dissolved in appropriate solvents at a concentration of 100 mg/ml for extracts and 10 mg/ml for isolates. 20 µl were dispensed on a 9 mm sterile paper disc (Munktel/Lasec, Numb. FLAS3526009). Amoxicillin was used as a positive control (40 µg/ml) for bacteria while Fluconazole (120 µg/ml) was used for fungi. The diluted cultures were spread on sterile Muller–Hinton agar plates. The plates were then incubated at 37°C for 18 to 24 h for bacterial pathogens and 3 days for fungal pathogens. The antimicrobial activity was evaluated by measuring the diameter of inhibition zone. The experiment was carried out in triplicate and the mean of the diameter of the inhibition zones was calculated. Antimicrobial inhibition activities measured were compared to that of Amoxicillin and Fluconazole standards for antifungal (Vlietinck, 1997).

DISCUSSION AND RESULTS

Aerial shoots and roots of *C. capensis* were used in this study. Extractions and isolations were specifically targeted for alkaloids and flavonoids indicated by the preliminary phytochemical screening results (Babajide et al., 2010) where high concentrations of flavonoid and alkaloids in the methanolic extracts of both the root and the aerial shoot were noted. A total of 6 compounds, viz- 1 to 6 were isolated from the fractionation of the TTA, methanolic extract of the aerial shoot and root. The TLC investigations of these 6 compounds revealed 1, 2 and 3 to be alkaloids as demonstrated by their positive test to dragendorff reagent using berberine as marker, while 4, 5 and 6 were flavonoids due to their positive test to vanillin in sulphuric acid using quercetin as a marker.

Compound 1 was obtained as a reddish-yellow powder with melting point of 85 to 87°C. The UV spectrum had a λ_{\max} values at 222 nm known to be associated with α,β -unsaturated carbonyl chromophore (Stuart, K.L., 1971) and that at 260 nm is typical for an aromatic ring system. The shoulder observed at 304 nm is a general characteristic of 8,14-dihydromorphinandienone alkaloids (Freitas et al., 1995). The IR spectrum showed pronounced bands at 3485 cm^{-1} (hydroxyl group), 2940 cm^{-1} (C-H of an olefinic group), 1685 cm^{-1} (α, β -unsaturated carbonyl), 1615 cm^{-1} (C=C olefinic) and 1500 cm^{-1} (C=C aromatic). The EI mass spectrum indicated the molecular formula to be $\text{C}_{20}\text{H}_{25}\text{NO}_6$ from the peak observed at M^+ (375). The following fragments were observed: 360 ($M - \text{CH}_3$)⁺, 332 ($360 - \text{CO}$)⁺, 238 ($M - \text{C}_8\text{H}_9\text{O}_2$)⁺ and 42 ($\text{H}_2\text{C}=\text{C}=\text{O}$)⁺, being characteristic of the morphinandienone alkaloids (Wheeler et al., 1968). The complete analysis of ¹H NMR, ¹³C NMR, HMBC, COSY and NOESY were used in assigning the structure of 1. The ¹H NMR (400 MHz, pyridine- *d*₅) showed only one aromatic signal at δ 6.91 which implies that the aromatic ring is penta-substituted. The four singlets integrated for three protons each were observed at δ 2.26 for (N-Me),

with the other three at δ 3.69 for MeO-6, 3.76 for MeO-3 and 3.83 for MeO-1. The two single proton signals at 10.75 and 11.50 were attributable to HO-4 and HO-5 respectively.

The ¹³CNMR (100MHz, pyridine-*d*₅) similarly showed a signal at δ 194.7 (s, C-7) for the α,β -unsaturated carbonyl, δ 43.1 (s, N-Me), δ 55.3 (s, C-6-OMe), δ 56.5 (s, C-3-OMe) and δ 57.2 (s, C-1-OMe). The complete assignments of all the protons and carbons are given in Table 1. This data was found to be in agreement with the literature (Blasko and Cordell, 1988; Vecchiotti et al., 1981) thus confirming compound 1 to be 5,6-dehydro-4,5-dihydroxy-1,3,6-trimethoxy- 17-methylmorphinan-7-one (Figure 1). Compound 1 to the best of our knowledge, has not been reported previously.

Compound 2 was obtained as a reddish-brown fluffy powder with m.p of 221 to 223°C. Its UV spectrum showed absorptions at 225, 243, 277 and 300 (shoulder) nm which are characteristic of 1, 2, 3-trisubstituted aporphinoid systems (Guinaudeau et al., 1979; Guinaudeau and Bruneton, 1993). The IR spectrum showed bands at 3420, 3050, 2925, 1636, 1458, 1387 and 1060 cm^{-1} confirming the aporphinic skeleton of 2. The EIMS showed an $[M]^+$ at 341.0112 which suggested the molecular formula of $\text{C}_{19}\text{H}_{19}\text{O}_5\text{N}$. ¹HNMR, ¹³CNMR, HMBC, COSY and NOESY spectra were used in assigning the structure. The ¹H NMR (400 MHz, $\text{C}_5\text{D}_5\text{N}$) spectrum showed an AB set of doublets at δ 5.94 and 6.13 ($J = 1.2$ Hz), characteristic of a methylenedioxy group at positions C-1 and C-2 in aporphines (Guinaudeau and Bruneton, 1993). The presence of an ABCD tetra substituted ring system with signals at δ 7.31 (br s) and 8.32 (br s) is a strong indication that ring D (the dimethoxysubstituted ring) is disubstituted. The ¹³C NMR (100 MHz, $\text{C}_5\text{D}_5\text{N}$) spectrum showed a signal at δ 101.3 (s, C-1, 2 methylenedioxy). In the ¹H - ¹H NOESY spectrum one of the methylenedioxy protons (δ 6.13) showed a correlation with H-11 (δ 8.32) while H-8 (δ 7.31) correlated with the H-7 methylene protons at (δ 2.95-3.10). Complete assignments of all protons and carbon atoms are listed in Table 2. The spectroscopic analysis above established the structure of 2 as 1, 2-methylenedioxy-3- hydroxy -9, 10-dimethoxyaporphine (Figure 2) and to the best of our knowledge it is being reported for the first time.

Compound 3 was isolated as a golden brown powder with m.p of 77 to 80°C and runs as a single spot in Hex: CHCl_3 : MeOH (3:10:2). This spot showed a characteristic orange colour of an alkaloid that is closely similar to the berberine standard TLC plate marker used when sprayed with dragendorff reagent. The IR spectrum showed a pronounced band at 3517 cm^{-1} due to the presence of a hydroxyl group while that at 2952 cm^{-1} was assigned to C-H of an olefinic system. The band at 1685 cm^{-1} is a characteristic of an α,β -unsaturated carbonyl system as observed for 1, the band at 1616 cm^{-1} is due to the olefinic C=C while that at 1510 cm^{-1} is due to the aromatic

Table 2. ^1H NMR (400 MHz) data of (**2**) and ^{13}C NMR (100 MHz) data of (**2**) in $\text{C}_5\text{D}_5\text{N}$.

C	δ ^{13}C	^1H
1	144.8	-
2	134.3	-
3	140.2	-
3a	118.0	-
4	25.3	2.92-3.12 2H m
5	44.1	2.92-3.12 1Hm, 3.45 1H m
6a	54.7	4.06 1H dd, (J = 5.2, 12.4 Hz)
7	37.1	2.95-3.10 2H m
7a	135.6	-
8	128.9	7.31 1H (br s)
9	160.2	-
10	161.9	-
11	127.1	8.32 1H (br s)
11a	133.7	-
11b	107.1	-
11c	130.2	-
O-CH ₂ -O	101.3	5.94, 6.13 2H A-B qJ = 1.2 Hz
MeO-9	56.5	3.76 s (3H)
MeO-10	55.3	3.69 s (3H)
HO-3	-	10.75 br s (1H)

C=C bond. Its UV spectrum showed absorptions at 226 nm which is known to be characteristic of an α,β -unsaturated carbonyl chromophore and at 261 nm due to an aromatic ring while the band at 302 nm is a characteristic of the 8,14-dihydromorphinandienone alkaloids as was noticed for **1** (Stuart, K.L., 1971). The EI mass spectral peak of 329.2133 suggested a molecular formula of $\text{C}_{19}\text{H}_{23}\text{O}_4\text{N}$ for **3**, for which the following fragments were observed: - (1) 314 $[\text{M} - \text{CH}_3]^+$, (2) 286 $[\text{314} - \text{CO}]^+$, (3) 192 $[\text{M} - \text{C}_8\text{H}_9\text{O}_2]^+$ and (4) 42 $[\text{H}_2\text{C}=\text{C}=\text{O}]^+$ confirming the characteristics of a morphinandienone alkaloid as observed for **1** (Wheeler et al., 1967). The ^1H NMR spectrum showed an AB pair of doublets at δ 6.90 and 6.78 (J = 8.4 Hz) for H-2 and H-1 respectively, while the three 3-proton singlets at δ 2.28 (N-Me), δ 3.62 (MeO-6) and δ 3.74 (MeO-3) (with similar numbering system as shown in **1**) were assigned as indicated. The singlet at δ 10.72 was attributable to HO-4. The ^{13}C NMR spectrum showed 19 carbon signals which supported the calculated molecular formula of $\text{C}_{19}\text{H}_{23}\text{O}_4\text{N}$. The carbons were assigned as shown in Table 3 where δ 43.3 is for N-Me, δ 55.2 for C6-OMe, δ 56.6 for C3-OMe and δ 194.5 for the C7-C=O. Complete assignments of all the protons and carbons are given in Table 3.

The structure of compound **3** was therefore established to be 5, 6-didehydro-4-hydroxy-3, 6-dimethoxy-17-methylmorphinan-7-one as shown in Figure 3. Literature reveals that **3** has been previously isolated from *Croton* (Vecchietti et al., 1981) and some *Cissampelos* species

such as *C. sympodialis* (Freitas et al., 1995) and *C. pareira* (Amresh et al., 2007). This is however the first time that the alkaloid has been reported as found in *C. capensis*.

Compound **4** was a yellow amorphous powder with a m.p. of 234 – 236°C. It showed EI mass spectral peak (M)⁺ at m/z 388.0103 suggesting a molecular formula of $\text{C}_{20}\text{H}_{20}\text{O}_8$. The IR spectrum showed pronounced bands at 3401 cm^{-1} (hydroxyl group), 2932 cm^{-1} (C-H olefinic), 1653 cm^{-1} (carbonyl group), 1612 cm^{-1} (C=C olefinic) and 1556 cm^{-1} (C=C aromatic). The UV spectrum showed two peak maxima at 357 and 255 nm respectively. The band at 271 nm showed a large bathochromic shift in the presence of sodium acetate, and an additional band appeared at 343 nm in the methanol NaOH spectrum which suggested a flavonol with a 7-OMe group. The ^1H NMR spectrum showed an aryl methyl singlet at δ 2.14 and four distinct methoxyl singlet peaks at δ 3.74 (OMe-3), 3.96 (OMe-3'), 4.01 (OMe-7) and 4.56 (OMe-8). The spectrum further showed the presence of only three 1-proton aromatic signals at δ 6.91 (d, J = 2.3 Hz), 7.60 (dd, J = 8.5 and 2.3 Hz) and 7.70 (d, J = 8.5 Hz) suggesting a 3',4'-disubstituted aromatic ring C, and a fully substituted aromatic ring A. The characteristically chelated 5-OH group was observed at δ 12.92 while that of 4'-OH was observed δ 10.90. By careful comparison of the ^{13}C NMR data of **4** with existing literature (Roitman and James, 1985; Breitmeier and Voelter, 1990; Babajide et al., 2008), the 4'-substituent was assigned to a hydroxyl

Table 3. ^1H NMR (400 MHz) data of (3) and ^{13}C NMR (100 MHz) data of (3) in $\text{C}_5\text{D}_5\text{N}$.

C	^1H	NOESY	HMBC
1	119.2	6.78 br d (J = 8.4 Hz)	C10, C12, C3
2	112.0	6.90 d (J = 8.4 Hz)	C1, C11, C4
3	147.2	-	-
3	145.6	-	-
4	125.3	-	-
5	8.30 s	-	C14, C12, C7 C6
6	152.2	-	-
7	194.5	-	-
8	40.5	ax 3.64 dd (J = 14.0, 17.6 Hz) eq. 2.60 dd (J = 17.6, 4.4 Hz)	C14, C7 C13
9	57.4	2.80 br d (J = 5.8 Hz)	C10, C14 C13, C14, C16, C11
10	28.4	ax 3.12 d (J = 17.2 Hz) eq 2.75 dd (J = 17.2, 5.8 Hz)	C9, C11 C14, C1, C12
11	132.2	-	-
12	128.2	-	-
13	38.5	-	-
14	42.0	2.40 m	-
15	33.2	2.22 - 2.12 m (2H)	C16 C12
16	47.6	2.42 m (1H)/2.30 m (1H)	C15 C13, C9
MeO-3	56.6	3.74 s (3H)	C3
MeO-6	55.2	3.62 s (3H)	C6
Me-N	43.3	2.28 s (3H)	C16, C9
HO-4	10.72 br s (1H)	-	-

hydroxyl group while that at the 3' position was assigned as a methoxyl group. Corroboration of this deduction was evident from chemical shifts of the substituted carbons, viz., C-3' at δ 147.2 and C-4' at 148.5. The other methoxyl groups were attached at C-3 (δ 137.5), C-7 (δ 145.4) and to C-8 (δ at 149.5 ppm which was in agreement with the HMBC correlations OMe-3/C-3 and OMe-7/C-7 and OMe-8/C-8 (Table 4).

The only non-oxygenated methyl group present was assigned as being attached to C-6, (which appeared at 106.6 ppm in the ^{13}C spectrum). Other peaks as demonstrated by the HMBC cross peaks were OH-5/C-5, C-6, C-4a; Me-6/C-5, C-6, C-7. Other carbon peaks were similarly assigned as shown in Table 4. Based on the above information the structure of 4 was suggested to be 3,7,8,3'-tetramethoxy-6-C-methyl-5,4'-dihydroxyflavone (6-C-methylquercetin 3, 3',7, 8 -tetramethyl ether) as shown in Figure 4 and to the best of our knowledge it is being reported for the first time.

Compound 5 was obtained as a yellow waxy powder with m.p of 185 to 187°C. A molecular formula of $\text{C}_{18}\text{H}_{18}\text{O}_9$ was suggested based on the molecular ion peak obtained at m/z 378.0625 and the fragmentation pattern in the EIMS. An exhaustive comparative study of the data obtained, revealed the general features of an isoflavanone nucleus (Mabry et al., 1970; Markham and Chari, 1982; Rahman and Gray, 2005; Rahman et al., 2007). The UV absorption spectrum with maxima at 333 and 288 nm suggested an isoflavanoid. The IR spectrum

had a strong absorbance at 3421 cm^{-1} due to an O-H group present, 2914 cm^{-1} for the C=C-H group, and 1634 cm^{-1} for the C=O system. The ^1H NMR spectrum showed an ABC pattern of 1-proton signals viz., δ 4.69 (*t*, $J = 11.0$ Hz), δ 4.50 (*dd*, $J = 11, 5.6$ Hz) and δ 4.43 (*dd*, $J = 11.0, 5.6$ Hz) assigned to the H-2 and H-3 protons respectively, which was similar to that observed in Markham and Chari's work as reported in 1982 for an isoflavanone system. The ^1H NMR spectral data (Table 5) also exhibited three H-bonded groups viz., C-5 hydroxyl at δ 12.89, C-7 hydroxyl at 12.98 and C-8 hydroxyl at δ 13.02. Two 1-proton singlets at δ 6.52 and δ 6.80 were assigned to H-6 of ring A and H-6' of ring C of the isoflavanone. The presence of 2 methoxyl groups as well as methylenedioxy group were apparent from their signals at δ 3.96 (2'-OMe), 3.87 (5'-OMe) and δ 5.90 (-OCH₂O-) respectively. The ^{13}C NMR spectrum demonstrated that compound 5 contained a total of 18 carbons including a carbonyl group as indicated by the IR absorption peak at 1634 cm^{-1} . The assignment of all carbons and the placement of the 2-methoxyl and methylenedioxy groups within the molecule were achieved by 2D NMR spectroscopic analysis. The HMBC data (Table 6) showed correlation of the C-5 hydroxyl group with a methine at δ 97.7 and a quaternary carbon at δ 103.3 which were assigned to C-6 and C-4a respectively.

The signals at δ 164.7, 168.6 and 169.3 were assigned to C-8a, C-7, and C-8 respectively as shown in Table 5.

Table 4. NMR spectroscopic data for compound (4).

¹³ C signal (δ)	DEPT	¹ H – ¹³ C COSY	HMBC	¹³ C Assignment
155.5	C	-	H-2', H-6'	2
137.5	C	-	3-Ome	3
178.8	-	-	-	4
156.3	C	-	-	5
106.6	C	-	-	6
145.4	C	-	7-Ome	7
149.5	C	-	C-7, C-9, C-10	8
151.4	C	-	8-Ome	8a
103.9	C	-	5-OH	4a
121.6	C	-	H- 5'	1'
115.3	CH	7.70d (J = 2.3 Hz)	H-6'	2
147.2	C	-	3'-OMe, H-5'	3'
148.5	-	-	H-5', H-6'	4'
115.8	CH	6.91d (J = 8.5 Hz)	-	5'
120.6	CH	7.60dd (J = 2.3 and 8.4 Hz)	H-2'	6'
59.5	CH ₃	3.74 s	C-5, C-6	3-Ome
60.2	CH ₃	4.01 s	C-7	7-Ome
63.5	CH ₃	4.56 s	C-8	8-Ome
56.5	CH ₃	3.96 s	C-3'	3'-Ome
8.1	CH ₃	2.14 s	-	6-Me

Correlation of the methoxyl protons with the carbons at δ 142.5 and 145.6 in the HMBC data proved the presence of C-2' and C-5' methoxy groups at these positions. The methylenedioxy group exhibited J₃ correlation with carbons at δ 137.8 and 149.8 and the occurrence of a J₃ correlation of H-6' to δ 149.8 suggested location of the methylenedioxy between C-3' and C-4'. The assignment of C-6' at δ 125.1 was also confirmed by the coupling observed in the HMQC which suggested the methine carbon at δ 48.5 as C-3 as observed in the J₃ for H-6'. Furthermore, the peak at δ 71.1 was assigned to C-2 as it showed a J₂ correlation to H-3 in the HMBC spectrum, and direct coupling with H-2 in the HMQC spectrum. The COSY spectrum showed correlation between H-3 and the two non-equivalent protons at C-2. The large diaxial coupling (J = 11.0 Hz) observed between H-3 and H-2 placed the aryl substituent at C-3 in the pseudo-equatorial position. Based on the above information the structure of compound 5 was therefore suggested to be 5,7,8-trihydroxy-2',5'-dimethoxy-3',4'-methylenedioxy-isoflavanone (Figure 5) and to the best of our knowledge it is being reported for the first time.

Compound 6 was isolated as a creamy yellow powder was assigned as C₁₇H₁₄O₈ based on the molecular ion peak at m/z 346.0752. The UV spectrum with maxima at 351 and 260 nm suggested a flavonol with a 7-OH group. A deep broad band observed in the IR spectrum at 3500 to 3400 cm⁻¹ was indicative of the presence of a hydrogen-bonded hydroxyl groups. The ¹H NMR spectrum exhibited two 3-proton singlets assignable to one

aryl methyl group at δ 2.11 (Me-6), very similar to that of 4 and one methoxyl group at δ 3.87 due to 3-OMe. The aromatic region showed three 1-proton signals typical of a 3', 4'-disubstituted ring C similar to that observed in 4 viz., δ 7.03 (d, J = 2.3 Hz), δ 7.62 (dd, J = 8.6 and 2.3 Hz) and δ 7.75 (d, J = 2.3 Hz). A broad cluster of 1-proton singlets at δ 10.23 (4'-OH), 11.15 (3'-OH), 12.05 (8-OH), 12.90 (7-OH) and 13.05 (5-OH) were due to five OH groups. The ¹³C spectrum (Table 7) demonstrated a pattern similar to that of compound 4, in which the C ring is di-substituted, carrying two hydroxyl groups at C-3' (163.6) and C-4'(164.5). The HMBC data for ring A was expected to be slightly different from that of 4 with the C-7(159.2) position showing the presence of an OH group. The HMBC cross peaks observed for OH-5/C-5, C-6, C-4a and Me-6/C-5, C-6, C-7, OH-8/C-7, C-8, C-8a indicated that the OH groups were located at C-7 and C-8 due to their respective coupling effects. The location of the single available methoxyl group and methyl group at C-3 and C-6 respectively, were further verified by their HMBC correlations with C-3 and C-6 respectively. Consequently the structure of 6 was with m.p of 217 to 219°C and the molecular formula of 6 assigned as 3-methoxy-6-C-methyl-3',4',5,7,8 -pentahydroxyflavone (6 - C- methylquercetin -3-methyl ether) (Figure 6). Compound 6 has, to the best of our knowledge, not been reported previously. It should be noted that there were several other extracts, fractions and subfractions that were not further analyzed due to insufficient quantities and time.

Table 5. ^1H NMR (400 MHz) data of (5) and ^{13}C NMR (100 MHz) data of (5) in $\text{C}_5\text{D}_5\text{N}$.

Position	^1H	^{13}C
2	4.69, <i>t</i> , $J = 11.0$ Hz	71.1
-	4.50, <i>dd</i> , $J = 10.5, 5.6$ Hz	-
3	4.43, <i>dd</i> , $J = 11.0, 5.6$ Hz	48.5
4	-	197.6
5	-	166.0
6	6.52, <i>s</i>	97.7
7	-	168.6
8	-	169.3
8a	-	164.7
4a	-	103.3
1'	-	121.4
2'	-	142.5
3'	-	137.8
4'	-	149.8
5'	-	145.6
6'	6.80, <i>s</i>	125.1
5-OH	12.89, <i>br. S</i>	-
7-OH	12.98, <i>br. S</i>	-
8-OH	13.02, <i>br. S</i>	-
2'-OMe	3.96, <i>s</i>	59.6
5'-OMe	3.87, <i>s</i>	60.2
-OCH ₂ O-	5.90, <i>s</i>	102.0

Table 6. HMBC data (400 MHz, $\text{C}_5\text{D}_5\text{N}$) of (5).

Proton	(5) ($^1\text{H} - ^{13}\text{C}$)	
	J_2	J_3
H-2	C-3	C-8a, C-4, C-1'
H-3	C-2, C-4, C-1'	C-2', C-6'
H-6	C-5, C-7	C-4a, C-8
H-6'	C-5'	C-3, C-2', C-4'
HO-5	C-5	C-6, C-4a
HO-7	C-7	C-6, C-8
HO-8	C-8	C-7, C-8a
MeO-2'	-	C-2'
MeO-5'	-	C-5'
-OCH ₂ O-	-	C-3', C-4'

Brine shrimp lethality bioassay

The brine shrimp lethality bioassay as described earlier was performed on all the extracts and fractions as shown in Babajide et al. (2010) and also on the isolates (1 to 6) estimating their LC_{50} with 95% confidence. As shown in Table 8, where three test bands were used viz., Inactive ($\text{LC}_{50} > 700$), active ($\text{LC}_{50} < 700$) and very active ($\text{LC}_{50} < 10$). The results demonstrated that all the hexane extracts showed no activity. Clearly evident was the finding that the highest bioactivity appeared in the

methanolic extracts, both in the root and the aerial shoot of this plant. The TTA showed a very high active value of 0.3155 (Table 8) thus indicating better activity than the BTA with a value of 64.87. This could be correlated with the isolation of the highly active compound 1 from the TTA. All the extracts and isolates (1, 3 and 5) that were very active should be considered as good candidates for antimicrobial and antiviral agents, since the lethality of a test substance to brine shrimp nauplii has been linked to the possible ability of such substances to kill cancer cells (antitumor activity) and possession of anti-inflammatory

Table 7. The NMR Spectroscopic Data for Compound (6).

¹³ C Signal (δ)	HMBC	¹ H (δ, J Hz)	¹³ C assignment
155.0	H-2', 6'	-	2
137.3	3-OMe	-	3
177.9	-	-	4
157.1	5-OH, Me-6	-	5
107.3	5-OH, Me-6	-	6
159.2	Me-6, 8-OH	-	7
162.8	-OH, Me-6	-	8
154.4	8-OH	-	8a
104.8	5-OH	-	4a
120.4	H- 5'	-	1'
115.4	H-6'	7.75 (1H, d, 2.3)	2'
163.6	H-2', H-5'	-	3'
164.5	H-2', H-5', H-6'	-	4'
115.4	-	7.03 (1H, d, 8.6)	5'
120.8	H-2'	7.62 (1H, dd, 2.3 and 8.6)	6'
7.2	-	2.11 (3H, s)	6-Me
59.2	-	3.87 (3H, s)	3-OMe

Table 8. Brine shrimp lethality test (average number dead counted after 24 hours).

Test materials	Vials at 1000 µg/ml	Vials at 100 µg/ml	Vials at 10 µg/ml	LC ₅₀	General remarks
CCRH	1	0	0	3526.22	Inactive
CCRM	10	10	9	3.75	Very active
-	-	-	-	-	-
CCAH	4	3	1	*	Inactive
CCAM	10	10	8	6.1523	Very active
-	-	-	-	-	-
TTA	10	10	9	0.3155	Very active
BTA	7	5	3	64.87	Active
-	-	-	-	-	-
1	10	10	10	0.8229	Very active
2	9	6	3	320.66	Active
3	10	9	7	6.201	Very active
4	8	6	3	199.55	Active
5	10	9	6	3.234	Very active
6	4	1	0	1159.62	Inactive

*Data did not converge and therefore could not be regressed by the finery probit analysis programme.

CCRH = *C. capensis* root nHex extract; CCRM = *C. capensis* root MeOH extract; CCAH = *C. capensis* aerial shoot nHex extract; CCAM = *C. capensis* aerial shoot MeOH extract; TTA = *C. capensis* aerial shoot nHexane extract; BTA = *C. capensis* aerial shoot neutralized aqueous residual fraction (Babajide et al., 2010)

properties (Mc Laughlin et al., 1991; Farthing, 2000). However, this does not mean that the inactive compounds (6 and hexane extracts) would not be useful for some other purposes such as a good antioxidant agents.

Antimicrobial activity evaluations

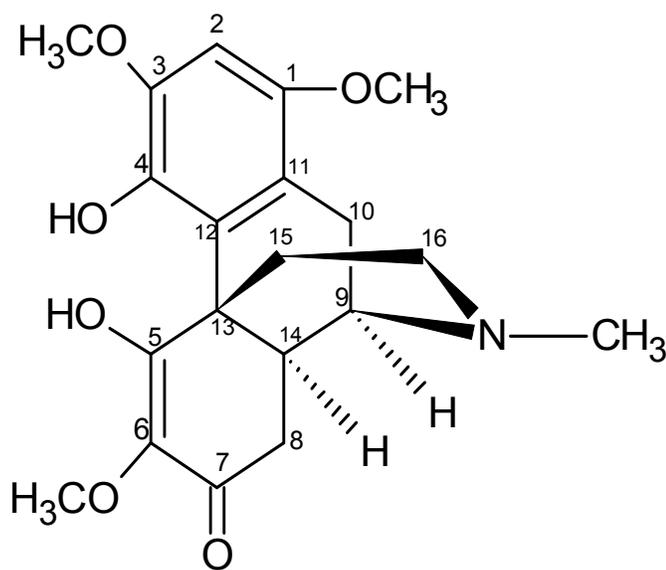
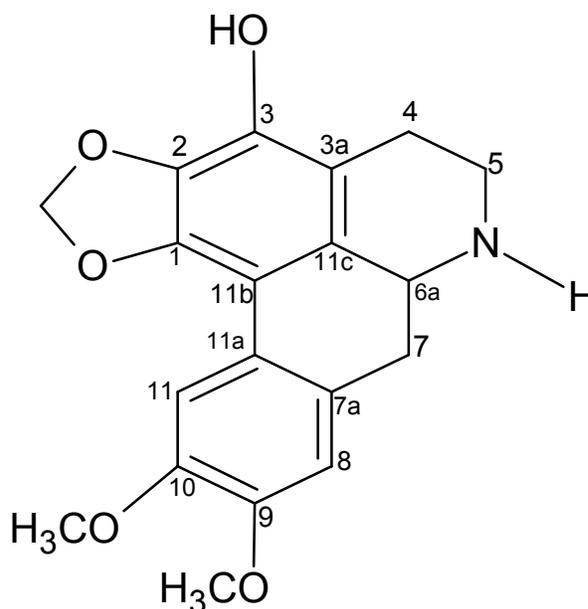
The antimicrobial evaluation was carried out using the

diffusion method in which three organisms were used in each case. For gram-negative bacteria: *P. aeruginosa*, *P. vulgaris* and *E. coli* Sero type 1 were used while for gram-positive, *B. subtilis*, *S. aureus* and *B. licheniformis* were used. The fungal species used were *C. albicans*, *C. eropiralis* and *A. niger*. The assay was set up as described earlier. The inhibition zones for both the bacteria and the fungi were measured in triplicate for all the extracts and the 6 identified isolates. The average was

Table 9. Antimicrobial profile of *Cissampelos capensis*.

Parameter	Psa	Prv	Esc	Bas	Sta	Bal	Caa	Cae	Asn
CCRH	0	0	0	0	0	0	0	0	0
CCRM	32	28	28	43	35	27	30	36	23
CCAH	0	0	0	0	0	0	0	0	0
CCAM	25	21	20	35	40	44	23	22	19
TTA	36	38	37	45	40	36	38	33	35
BTA	15	17	17	23	28	24	30	32	29
1	41	34	29	32	30	33	0	0	0
2	16	19	19	22	19	14	19	15	17
3	18	0	26	19	0	18	10	11	11
4	10	13	11	12	10	11	0	0	14
5	15	13	19	22	28	30	35	29	27
6	10	11	15	0	0	21	20	26	29
Amx	61	44	49	42	53	47	0	0	0
Flu	0	0	0	0	0	0	46	53	39

*Gram -ve bacteria: - *Pseudomonas aeruginosa* (Psa), *Proteus vulgaris* (Prv) and *Escherichia coli* (Esc) Gram +ve bacteria: - *Bacillus subtilis* (Bas), *Staphylococcus aureus* (Sta) and *Bacillus licheniformis* (Bal) Fungi: - *Candida albicans* (Caa), *Candida eropiralis* (Cae) and *Aspergillus niger* (Asn) Amx:- Amoxicillin and Flu:- Fluconazole.

**Figure 1.** (1) 5, 6-dehydro-4,5-dihydroxy-1,3,6-trimethoxy-17-methylmorphinan-7-one.**Figure 2.** (2) 1,2-methylenedioxy-3-hydroxy-9,10-dimethoxyaporphine.

determined and compared to the positive controls used (Amoxicillin; 40µg/ml for bacteria and Fluconazole; 120µg/ml for fungi). Similar patterns were observed in the microbial analysis except in very few cases as shown in Table 9. The selection of the three (3) sets of test organisms was designed to allow for the study of a reasonably wide spectrum of antimicrobial activities of the extracts and isolates.

The results showed different levels of activity for both the extracts and the isolates. Any value > 20 mm is

assumed to be moderately active. There was more antibacterial than antifungal activity. The highest activity was recorded for the gram +ve bacteria while that of gram -ve bacteria was more than the fungi. The maximum zone (45 mm) of antibacterial activity was observed in the TTA fraction against the gram +ve organism *B. subtilis*. This may be due to the strong antimicrobial and antiviral activities of the alkaloids in the plant that is well known for the treatment of malaria, fungal infections,

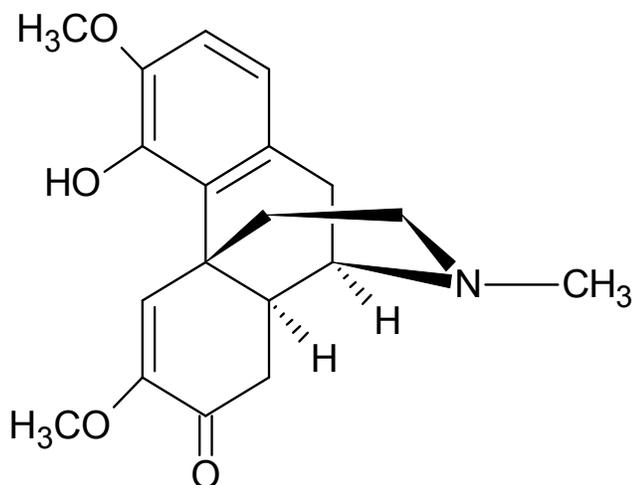


Figure 3. (3) 5, 6-didehydro-4-hydroxy-3, 6-dimethoxy-17-methylmorphinan-7-one.

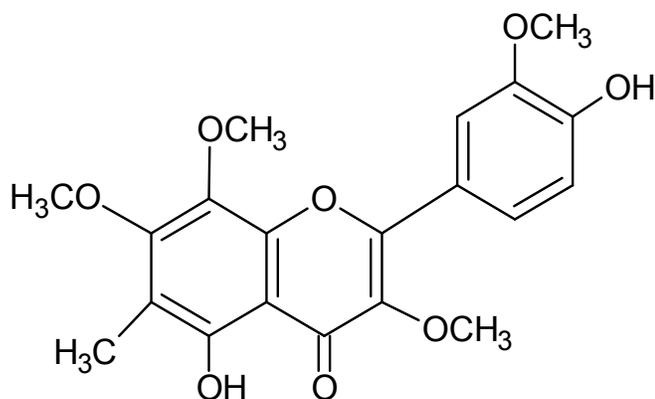


Figure 4. (4) 6-C-methylquercetin 3, 3', 7, 8-tetramethyl ether.

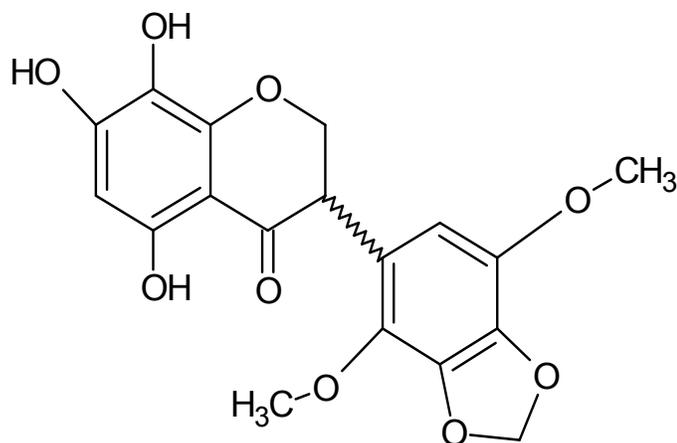


Figure 5. (5) 5, 7, 8-Trihydroxy-2'5'-methoxy-3', 4'-methylenedioxyisoflavanone.

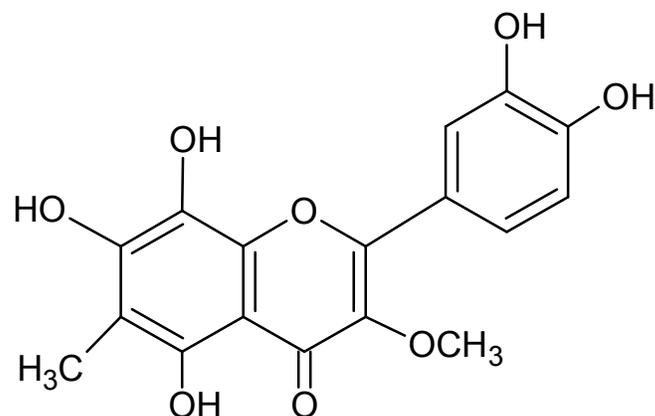


Figure 6. (6) 6-C-Methylquercetin 3-Methyl ether.

inflammations and cancer (Kaur et al., 2009; Mc Gaw et al., 2000). No activity was recorded for the hexane extracts against all nine organisms. High values of activity were recorded for the drug resistant breed of bacteria *P. aeruginosa*, where an inhibition zone of 36 mm was recorded for TTA, 25 mm for CCAM and 32 mm for CCRM.

Some preferential selective activity was observed in which the compound preferentially act on certain selected organism were inactive against the other set (Babajide et al., 2010; Shai et al., 2008). Similar observations were made in the isolated compounds, where high values were recorded for 1 against gram +ve and -ve bacteria whereas there was no effect on all the three fungi. High fungicidal effect was recorded for 5 while the activities were generally low for compound 4. The observed biological activities of extracts from this plant may thus be due to the presence of both alkaloids and flavonoids, as these compounds are generally known to exhibit a wide range of activities which include anti inflammatory, antithrombotic, anticancer and antiviral, where some of these may be associated with their ability to scavenge free-radicals (Babajide et al., 2008; Shai et al., 2008; Rahman et al., 2007; Liu et al., 1990).

Conclusion

This study successfully relates the specific constituents present in *C. capensis* to its widespread ethnomedicinal uses. The results provide a useful profile of active compounds for the development of useful and promising anticancer, antimicrobial and antioxidant agents in the future.

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Conflict of interest

The authors declare that they have no conflict of interest.

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