

A global infrageneric classification system for the genus *Crotalaria* (Leguminosae) based on molecular and morphological evidence

Marianne M. le Roux, James S. Boatwright and Ben-Erik van Wyk

Abstract

Crotalaria is a large genus of 702 species with its centre of diversity in tropical Africa and Madagascar and secondary radiations in other parts of the world. The current infrageneric classification system is based on morphological and morphometric studies of the African taxa only and is here re-evaluated using a phylogenetic approach. DNA sequences derived from the nuclear ITS and the plastid *matK*, *psbA-trnH* and *rbcLa* markers were analyzed using parsimony and model-based (Bayesian) approaches. The resultant molecular phylogeny allowed for a new interpretation of diagnostically important morphological characters, including specialisations of the calyx, keel, standard petal and style, which are variously convergent in several unrelated infrageneric groups. Of particular interest is the congruence between the new phylogeny and the distribution of standard petal callosity types. A sectional classification system for the entire genus is proposed for the first time. The new system that is formalised here comprises eleven sections: *Amphitrichae*, *Calycinae*, *Crotalaria*, *Geniculatae*, *Glaucæ*, *Grandifloræ*, *Hedriocarpæ*, *Incanæ*, *Schizostigma*, *Borealigeniculatae* and *Stipulosæ*. Sectional limits of the *Geniculatae*, *Calycinae* and *Crotalaria* are modified. The subsections *Stipulosæ*, *Glaucæ* and *Incanæ* are raised to sectional level, while some groups previously recognized as subsections are abandoned due to non-monophyly (subsections *Chrysocalycinae*, *Hedriocarpæ*, *Macrostachyæ* and *Tetralobocalyx*). Two new sections are recognized, *Amphitrichae* and *Borealigeniculatae*.

Introduction

The large genus *Crotalaria* L. comprises 702 species and is widely distributed, especially in the Southern Hemisphere (Van Wyk, 2005; Jianqiang & al., 2010; Le Roux & al., subm.). It has its primary centre of species diversity in tropical and subtropical Africa and Madagascar (Polhill, 1982; Van Wyk, 2005; Flores & al., 2006), where some 543 species occur, and secondary radiations in temperate Asia, tropical Asia and Australasia with 159 species, South America (especially Brazil) with 64 species and North America (Mexico and the southern parts of the United States of America) with 34 species (Le Roux & al., subm.; Flores & al., 2006; Pandey & al., 2010). These figures include the total number of species in each individual region, i.e., species that occur in more than one region are included in each region.

The genus is part of the largely African tribe Crotalariaeae, and together with *Bolusia* Benth. and *Euchlora* Eckl. & Zeyh. sister (100% BS, PP 1.0; Boatwright & al., 2008a) to the remainder of the tribe (Polhill, 1976, 1982; Van Wyk, 1991, 2003, 2005; Van Wyk & Schutte, 1989, 1995; Boatwright & al., 2008a, 2011). *Crotalaria* is easily recognized by a unique combination of characters, namely a rostrate keel, highly inflated fruit, a hairy style, a 5+5-anther configuration, paired callosities on the standard petal and the presence of macrocyclic pyrrolizidine alkaloids (Polhill, 1982; Van Wyk & Verdoorn, 1990; Van Wyk, 2005). *Bolusia* differs from *Crotalaria* in having a helically coiled keel with a single callosity restricted to the standard petal blade (Van Wyk & al., 2010; Le Roux & Van Wyk, 2012). *Euchlora* lacks standard petal callosities and has an obtuse to somewhat rostrate keel beak (Le Roux & Van Wyk, 2012).

Polhill (1968) re-evaluated the infrageneric classification system following previous work by Bentham (1843), Harvey (1862), Baker (1914), Verdoorn (1928), Wilczek (1953a, b), Hepper (1958), Milne-Redhead (1961, and unpub.), Torre (1962) and Schreiber (1970). He recognized eleven sections and eight subsections in Africa and Madagascar, mainly derived from floral and fruit characters. Morphometric studies were also conducted by Bisby (1970, 1973), who wanted to illustrate the application and potential value of numerical taxonomic methods and used *Crotalaria* as model genus. Bisby (1973) noted the difficulty of classifying *Crotalaria* into natural groups of workable sizes, due to the large number of species and high levels of interspecific variation potentially caused by reticulation.

Table 1. The infrageneric classification systems for the genus *Crotalaria* since 1973.

	Ansari (2006, 2008)	Le Roux & al. (proposed here)
Bisby & Polhill (1973), Polhill (1982)		
Sect. 1: <i>Grandiflorae</i> (Baker f.) Polhill	Sect. 1: <i>Calycinae</i> Wight & Arn.	Sect. 1: <i>Hedriocarpae</i> Wight & Arn. emend. M.M. le Roux & B.-E. van Wyk
Sect. 2: <i>Chrysocalycinae</i> (Benth.) Baker f.	Subsect. <i>Alatae</i> (Wight & Arn.) A.A. Ansari,	Sect. 2: <i>Incanae</i> (Benth.) Polhill
Subsect. <i>Chrysocalycinae</i>	Subsect. <i>Calycinae</i>	Sect. 3: <i>Schizostigma</i> Polhill
Subsect. <i>Glaucae</i> (Benth.) Bisby & Polhill	Subsect. <i>Diffusae</i> (Wight & Arn.) A.A. Ansari	Sect. 4: <i>Calycinae</i> Wight & Arn. emend. M.M. le Roux & B.-E. van Wyk
Subsect. <i>Incanae</i> (Benth.) Bisby & Polhill	Subsect. <i>Heylandiae</i> A.A. Ansari	Sect. 5: <i>Borealigenticulatae</i> M.M. le Roux & B.-E. van Wyk
Subsect. <i>Stipulosae</i> (Baker f.) Bisby & Polhill	Sect. 2: <i>Chrysocalycinae</i> (Benth.) Baker f.	Sect. 6: <i>Crotalaria</i> emend.
Subsect. <i>Tetralobocalyx</i> (Harms.) Bisby & Polhill	Subsect. <i>Incanae</i> (Benth.) Bisby & Polhill	Sect. 7: <i>Stipulosae</i> (Baker f.) M.M. le Roux & B.-E. van Wyk
Sect. 3: <i>Hedriocarpae</i> Wight & Arn.	Subsect. <i>Stipulosae</i> (Baker f.) Bisby & Polhill	Sect. 8: <i>Glaucae</i> (Benth.) M.M. le Roux & B.-E. van Wyk
Subsect. <i>Hedriocarpae</i>	Sect. 3: <i>Crotalaria</i>	Sect. 9: <i>Genticulatae</i> Polhill
Subsect. <i>Macrostachyae</i> (Benth.) Bisby & Polhill	Subsect. <i>Bracteatae</i> (Wight & Arn.) A.A. Ansari	Sect. 10: <i>Amphitrichae</i> M.M. le Roux & B.-E. van Wyk
Sect. 4: <i>Genticulatae</i> Polhill	Subsect. <i>Crotalaria</i>	Sect. 11: <i>Grandiflorae</i> (Baker f.) Polhill
Sect. 5: <i>Schizostigma</i> Polhill	Subsect. <i>Longirostrae</i> (Benth.) Polhill	
Sect. 6: <i>Calycinae</i> Wight & Arn.	Subsect. <i>Polyphyllae</i> (Wight & Arn.) A.A. Ansari	
Sect. 7: <i>Crotalaria</i>	Sect. 4: <i>Dispermae</i> Wight & Arn.	
Subsect. <i>Crotalaria</i>	Sect. 5: <i>Grandiflorae</i> (Baker f.) Polhill	
Subsect. <i>Longirostris</i> (Benth.) Polhill	Sect. 6: <i>Hedriocarpae</i> Wight & Arn.	
Sect. 8: <i>Dispermae</i> Wight & Arn.	Subsect. <i>Hedriocarpae</i>	
	Subsect. <i>Macrostachyae</i> (Benth.) Bisby & Polhill	

In 1973, Polhill & Bisby combined their datasets which resulted in small changes to the system proposed by Polhill (1968), with eight sections and nine subsections (listed in Table 1) recognized in the revision of the African and Madagascan species (Polhill, 1982). The new circumscriptions were mainly based on floral characters.

Polhill distinguished between two groups in *Crotalaria*, derived from the distribution of callosities associated with the standard petal. They are referred to by Polhill (1982) as the “unspecialised group” which includes sections *Grandiflorae*, *Chrysocalycinae* and *Hedriocarpae* while the so-called “specialised group” includes sections *Calycinae*, *Crotalaria* and *Dispermae* and two “intermediate sections”, *Geniculatae* and *Schizostigma* (Bisby & Polhill, 1973). The term “specialised” refers to those taxa that exhibit a whole suite of morphological adaptations that are related to an increasing level of complexity in the pollination syndrome. Unspecialised floral characters include callosities present on the standard petal blade and claw (Fig. 1A1), a rounded keel with the curvature at about or below the middle (Fig. 1B1, B2), a straight keel beak (rarely circumflexed or twisted in older flowers) and a rounded style (Fig. 1F7), usually with one line of trichomes (Fig. 1F1, F4). Specialised floral characters include callosities restricted to the blade (Fig. 1A2–A4), a rounded, subangled or angled keel (Fig. 1B5, B6) with the curvature rarely at about the middle, usually below the middle or angled in the lower third, a twisted keel beak (except in section *Geniculatae*) and a rounded (Fig. 1F7) or geniculate (Fig. 1F8) style with two lines of trichomes (Fig. 1F2, F5) (or less frequently one line; Le Roux & Van Wyk, 2012).

Ansari (2006, 2008) published a revision of the infrageneric classification system of *Crotalaria* to accommodate the Indian species. Six of the eight sections as recognized by Polhill (1982) (Table 1) are present in India. Ansari created four new subsections in section *Calycinae* and two new subsections in section *Crotalaria* using floral and vegetative characters, but he did not consider the total range of variation of these characters across the genus (extra-Indian species). Apart from studies at the generic level (Boatwright & al., 2008a), no molecular systematic studies of the large and unwieldy genus *Crotalaria* have yet been attempted. Therefore this study was aimed at presenting a first molecular phylogeny for the genus based on nrDNA internal transcribed spacer (ITS) sequences, along with plastid *matK*, *psbA-trnH* and *rbcLa* sequences. The phylogenetic approach is used to identify monophyletic groups, to assess the congruence between morphological and sequence-based patterns and to evaluate the current hypotheses of infrageneric relationships based on regional Floras (Polhill, 1982; Ansari, 2006, 2008). The ultimate aim of our study was not to merely explore monophyly, but to propose, for the first time, a practical and workable global infrageneric classification system, based on careful consideration of the main patterns of character state distributions in all species from all continents. We were also interested in exploring new ways of interpreting the evolution of salient morphological features of the flowers (Le Roux & Van Wyk, 2012) that were previously used by Polhill (1982) as diagnostic characters to define sections and subsections.

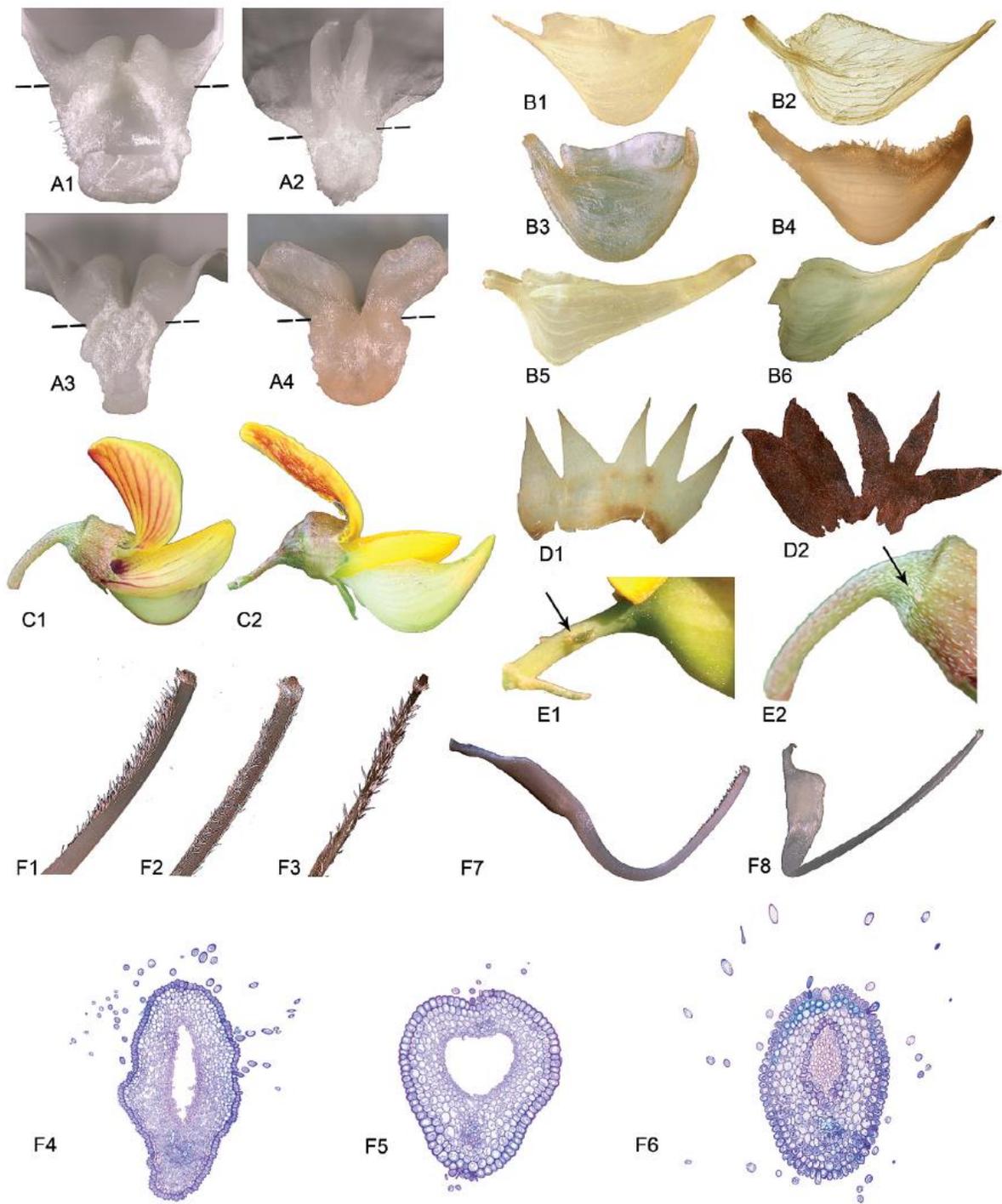


Fig. 1. Diagnostic floral characters and character states in *Crotalaria*. **A**, shape and distribution of callosities on the standard petal; **B**, shape and vestiture of the keel petals and shape of the beaks; **C**, **D**, shape of the calyx tubes and lobes; **E**, position of the bracteoles on the pedicel; **F**, vestiture and shape of styles. **A1**, ridge callosities present on the blade and claw; **A2**, columnar callosities present on the blade only; **A3**, disc callosities present on the blade only; **A4**, lamelliform callosities present on the blade only (the dotted line indicates the division between the standard petal blade and claw); **B1**, a rounded keel with an untwisted beak; **B2**, a sub-angled keel with a twisted beak; **B3**, a rounded keel with a circumflexed beak; **B4**, a rounded keel with an untwisted beak, but with lanate-pilose trichomes on the upper edge of the keel; **B5**, angled keel with an untwisted beak; **B6**, angled keel with a twisted beak; **C1**, truncate calyx tube; **C2**, campanulate calyx tube; **D1**, sub-equally lobed calyx; **D2**, bilabiate calyx; **E1**, bracteoles inserted on the pedicel; **E2**, bracteoles inserted on the calyx; **F1**, **F4**, style in side view and cross section, with trichomes in a single line along the upper edge; **F2**, **F5**, style in side view and cross section, with trichomes in two lines along the upper and lower edges; **F3**, **F6**, style in side view and cross section, with dispersed trichomes all around the style; **F7**, rounded style; and **F8**, geniculate style.

Materials and methods

Taxon sampling. — Fieldwork was conducted in Botswana, Ethiopia, Namibia and South Africa to collect silica-dried leaf material for DNA sequencing. Additional samples were either obtained as gifts or were taken from herbarium specimens to include representative samples of all sections in the current infrageneric classification system from all continents. Voucher specimen information is listed in Appendix S1 along with author citations for all taxa and these are not repeated elsewhere. A sister relationship was shown between *Bolusia* and *Crotalaria* (Polhill, 1976, 1982), which was recently confirmed by molecular studies (92% BS, PP 1.00; Boatwright & al., 2008a, 2011). Furthermore, the latter studies indicated that *Crotalaria*, *Bolusia* and the monotypic genus *Euchlora* form a strong clade (99% BS, PP 1.00), therefore both *Euchlora* and *Bolusia* were chosen as outgroups in this study (Boatwright & al., 2008a).

DNA extraction, amplification and sequencing. — The majority of DNA extractions were carried out at the Molecular Systematics Laboratory at the University of Johannesburg (UJ), South Africa. Two extraction methods were used: (1) silica-dried leaf material (0.1–0.3 g) was extracted using the 2× hexadecyltrimethylammonium bromide (CTAB) method (Doyle & Doyle, 1987) and purified using QIAquick silica columns (Qiagen Inc., Hilden, Germany); (2) leaf material sampled from herbarium specimens (0.008–0.020 g) was extracted and purified with a DNeasy plant minikit (Qiagen Inc.) following the manufacturer's protocol. A total of 15 DNA samples were obtained from Kew Herbarium (K), and were extracted by the Jodrell Laboratory, U.K. The DNA regions were amplified using polymerase chain reactions (PCR) in 25 µl reactions, following Boatwright & al. (2008a), but with 10–40 ng DNA template, 1 µl 0.004% bovine serum albumin (BSA) and 1% dimethyl sulfoxide (DMSO) in the PCR reactions for amplification of ITS. Successfully amplified PCR products were purified using either the QIAquick PCR purification kit (Qiagen Inc.), according to the manufacturer's instructions, or the ExoSAP protocol of Werle & al. (1994) using 5 units of Exonuclease I and 0.5 units of Shrimp Alkaline Phosphatase. Four DNA regions were amplified: nuclear ribosomal ITS (primers of White & al., 1990 and Sun & al., 1994); barcoding *matK* (standard barcoding primers of Ki-Joong Kim: 3F_KIM f: CGT ACA GTA CTT TTG TGT TTA CGA G; 1R_KIM r: ACC CAG TCC ATC TGG AAA TCT TGG TTC); barcoding *rbcLa* (standard barcoding primers of Kress & Erickson, 2007) and; *psbA-trnH* (primers of Sang & al., 1997).

The ITS region was chosen because previous studies by Boatwright & al. (2008a, 2011) have shown that it provides robust resolution at higher taxonomic levels for the tribe Crotalariaeae. Three additional regions were chosen, one that is phylogenetically conservative (*rbcLa*) and two that are evolving more rapidly, *matK* and *psbA-trnH*. These regions have been under consideration as DNA barcoding markers and have shown to be informative (Kress & Erickson, 2007; Edwards & al., 2008; Lahaye & al., 2008; Kress & al., 2009).

Sequences were completed for the barcoding plastid gene regions *rbcLa* and *matK* at the UJ lab as well as the Canadian Centre for DNA Barcoding (University of Guelph, Canada; see Appendix S1) using standard barcoding PCR and cycle sequencing programs and loaded onto the Barcode of Life Data System (BOLD; Ratnasingham & Hebert, 2007). The ITS and *psbA-trnH*

regions were solely amplified at UJ using PCR and cycle sequencing programs as described by Boatwright & al. (2008b) and Downie & Katz-Downie (1996) in problematic cases (where degraded DNA was extracted from old herbarium specimens and as a result were difficult to amplify).

Sequence alignment and phylogenetic analyses. — Complementary sequences were assembled and edited in Sequencher v.4 (Gene Codes Corporation), aligned manually in PAUP* v.4.0b10 (Swofford, 2002) and in MEGA v.4 (Tamura & al., 2007). No problematic areas were encountered during the alignment of the datasets and particular care was taken in the case of *psbA-trnH* following reports of frequent inversions in the region that might affect estimation of relationships (Bain & Jansen, 2006; Whitlock & al., 2010). Insertions and deletions were coded as missing data and did not contribute to the analyses. The DNA sequences have been submitted to GenBank (accession numbers listed in Appendix S1) and the datasets to TreeBASE (submission number 12059).

Parsimony analysis (MP; Fitch, 1971) and Bayesian Inference (BI) were used to analyse the data. MP analyses were performed in PAUP* for the four individual gene matrices using a heuristic search with 1000 random sequence additions and holding 10 trees per replicate with tree bisection-reconnection (TBR) branch swapping and the MULTREES options in effect. Delayed transformation character optimization (DELTRAN) was used to illustrate branch lengths. Internal support was determined using bootstrap analysis (Felsenstein, 1985) with 1000 replicates holding 10 trees per replicate, TBR branch swapping and MULTREES options in effect. Bootstrap values were evaluated according to the following scale: 50%–74%, low; 75%–84%, moderate; 85%–100%, strong. The bootstrap consensus trees for each of the four individual datasets were evaluated for congruence. We relied only on visual inspection of the bootstrap consensus trees and no statistical analysis were done due to the unreliability of tests such as the length incongruence test (ILD; Farris & al., 1995) shown in previous studies (Reeves & al., 2001; Yoder & al., 2001). In a phylogenetic study of the tribe Crotalariaeae, Boatwright & al. (2008a) also found that in the absence of strongly supported incongruent patterns between datasets, the ILD test indicated significant incongruence and was unreliable to detect incongruent patterns compared to visual inspection. Their suggestion and that of Seelanan (1997) is therefore followed and datasets combined if no strongly supported incongruent patterns (85%–100% BS or “hard” incongruence) were detected.

No incongruent patterns were noticed between any of the datasets. Two combined analyses were conducted: (1) the ITS and *rbcLa* combined analysis (dataset 1; 186 accessions, 168 species); and (2) the ITS, *matK*, *psbA-trnH* and *rbcLa* combined analysis (dataset 2; 141 accessions, 131 species). These analyses included representatives of all major groups as classified by Polhill (1982) and most of the subsections (dataset 1) without missing data.

For BI (Huelsenbeck & Ronquist, 2001; Ronquist & Huelsenbeck, 2003) a partitioned analysis was implemented and the GTR+I+G model selected for both the ITS and *rbcLa* datasets, the TVM+G model for the *matK* dataset, and the GRT+G model for the *psbA-trnH* dataset

indicated by ModelTest v.3.1.2 (Posada & Crandall, 1998) using the Akaike information criterion (AIC). The BI analyses were run in MrBayes v.3.1.2 for a total of 16 million generations sampling every 10,000 generations for dataset 1 and 11 million generations sampling every 10,000 generations for dataset 2. Tracer v.1.5.0 (Rambaut & Drummond, 2009) was used to assess convergence, comparing traces of two simultaneous analyses, determining whether adequate sample sizes (ESS) were used for each parameter, if the MCMC parameter samples were taken from a stationary, unimodal distribution, and whether stationarity of posterior probabilities of splits within runs were reached. After the analyses were completed, 25% of the trees were removed as the “burn-in” and majority-rule consensus trees were produced for each of the two analyses. Posterior probability (PP) values (only those above 0.5 are indicated) were evaluated according to the following scale: 0.50–0.94, weak; 0.95–1.00, strong.

Results

Statistics for each dataset obtained in the parsimony analyses are listed in Table 2.

Combined ITS and *rbcLa* dataset. — The bootstrap consensus trees from the individual ITS and *rbcLa* datasets were compared and no evidence of strongly supported incongruent patterns was found. These two datasets were combined into a single analysis, because it contained the largest number of species for which both regions were successfully sequenced and resulted in a well-resolved phylogeny at the terminal clades. *Crotalaria* is monophyletic (99% BS, PP 1.0) with well-resolved clades corresponding to some of Polhill’s (1982) infrageneric groups (Fig. 2).

Combined ITS, *matK*, *psbA-trnH* and *rbcLa* dataset. — Both the MP and BI analyses showed similar results in the total combined analysis (Fig. 3) and indicated that *Crotalaria* is monophyletic (99% BS, PP 1.0). The phylogeny is fairly well-resolved at the branch ends, with a total of 15 clades that were identified, but with an unresolved backbone. Nine of these clades correspond to the infrageneric concepts as proposed by Polhill (1982) as either sections or subsections.

Discussion

The molecular phylogeny (Figs. 2, 3) reveals eleven monophyletic groups that are variously supported by bootstrap percentages, Bayesian posterior probabilities and morphological evidence. Monophyletic groups are supported by combinations of morphological characters rather than single, unambiguous synapomorphies. These unique combinations are listed in Table 3 and are briefly discussed for each clade:

1. Hedriocarpace clade (PP 0.56). All species have ridge callosities present on the standard petal blade and claw, a rounded (rarely angled) keel with an untwisted beak and a rounded (rarely geniculate) style with trichomes in one line. Most species have a truncate calyx (sometimes only slightly so) with bracteoles inserted at the base or just below the calyx. The five calyx lobes are shorter than the keel and more or less subequally lobed or as long as the keel and slightly bilabiate. Petals often have purple or brown-coloured venation.

2. Stipulosae clade (77% BS, PP 1.00). Asymmetrical stipules are present in most of the species. Ridge callosities are invariably present on the standard petal blade and claw. The keel is somewhat rounded or subangled at about or below the middle to angled in the lower third with an untwisted beak (rarely incurved, becoming slightly coiled) and lanate hairs along the upper edge. The fruit is usually cylindrical or oblong-ellipsoid.

3. Genuculatae clade (72% BS, PP 0.96). Columnar or disc callosities are restricted to the standard petal blade. The keel is angled in the lower third with an untwisted beak and a geniculate style with one (those with disc callosities) or two (those with columnar callosities) lines of hairs (amphistylar hairs present in *C. monophylla*). The fruit is thick-walled (if somewhat thin-walled, then inflorescences with twelve or more flowers).

Table 2. Statistics obtained for the parsimony analyses for each of the four individual regions sequenced (ITS, *matK*, *psbA-trnH*, *rbcLa*) as well as the combined ITS and *rbcLa* regions (dataset 1) and combined ITS, *matK*, *psbA-trnH* and *rbcLa* regions (dataset 2).

	ITS	<i>matK</i>	<i>psbA-trnH</i>	<i>rbcLa</i>	Dataset 1 (ITS, <i>rbcLa</i>)	Dataset 2 (ITS, <i>matK</i> , <i>psbA-trnH</i> , <i>rbcLa</i>)
No. of included accessions	235	161	211	198	186	141
No. of included taxa	207 (30%)	145 (21%)	190 (27%)	178 (25%)	168 (24%)	131 (19%)
No. of included characters	818	720	610	552	1355	2671
No. of variable characters	588 (72%)	214 (30%)	291 (48%)	95 (17%)	462 (34%)	832 (31%)
No. of parsimony-informative sites	404 (49%)	103 (14%)	166 (27%)	64 (12%)	304 (22%)	474 (18%)
No. of trees (Fitch)	1920	9661	80	2440	5180	6450
No. of steps (tree length)	2419	386	1960	455	1552	2059
Consistency index (CI)	0.39	0.67	0.22	0.25	0.43	0.54
Retention index (RI)	0.70	0.87	0.21	0.31	0.80	0.80

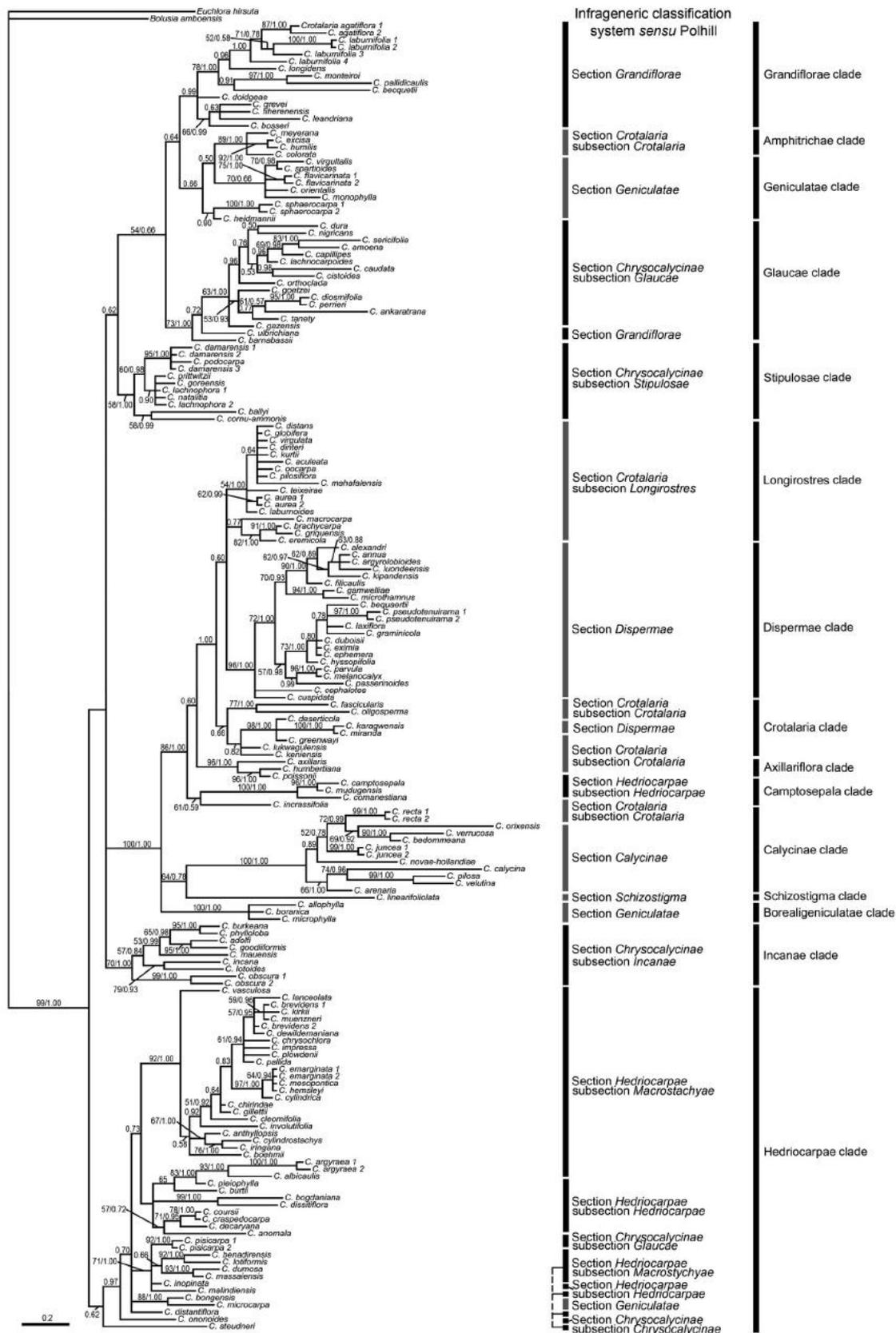


Fig. 2. The combined ITS and *rbcLa* analysis (dataset 1) of 168 species (186 accessions) produced similar trees in the Bayesian and maximum parsimony (tree length = 1552, consistency index = 0.43, retention index = 0.80) analyses. A 50% majority-rule phylogram from the BI is illustrated with bootstrap percentages above 50% indicated left of the forward slash and posterior probability values above 0.5 right of the forward slash. A total of 15 clades were identified and compared with the current classification system proposed by Polhill (1982). Sections forming part of Polhill's "specialised" group are shown in grey and sections included in the "unspecialised" group are shown in black. The number of expected substitutions per site is indicated by the scale.

Table 3. A comparison of morphological diagnostic characters (brackets indicate rare character states) for each of the eleven recognized infrageneric groups of the genus *Crotalaria*.

Character	<i>Hedriocarpae</i>	<i>Incanae</i>	<i>Schizostigma</i>	<i>Calycinae</i>	<i>Boreali-geniculatae</i>	<i>Crotalaria</i>	<i>Stipulosae</i>	<i>Glaucæ</i>	<i>Geniculatae</i>	<i>Amphirichae</i>	<i>Grandiflorae</i>
Habit (- tree, + shrub, ++ perennial, +++ annual)	++/ +++	+ +++	(+) +++	(+) +++	+/	(+) +++	(+) (+++)	+ (+++)	(+) +++	+/	-/+
Leaves (- simple, + compound)	+	+	+	-	+	(-)	+	-	(-)	+	(-)
Stipule symmetry (- absent or inapplicable, + symmetrical, ++ asymmetrical)	+	+	+	+/	-	-	(-)	-	-	+	-
Stipule shape (- not decurrent, + decurrent)	-	-	-	-	-	-	-	-	-	-	-
Bracteole position (- absent or inapplicable, + on or below calyx, ++ on pedicel)	+	+	-	+/	++	+/	++	(+)	++	++	++
Calyx type (- 5-subequally lobed, + 4-subequally lobed, ++ bilabiate)	-	-	-	(-)	-	-	-	-	-	-	-
Calyx tube (- campanulate, + truncate)	(-)	-	-	-	-	-	-	-	-	-	-
Calyx length (- shorter than keel, + as long as keel, ++ longer than keel)	-	+	+	(-)	-	(-)	(-)	(-)	-	(-)	-
Callosity shape (+ ridge, ++ disc, +++ lamelliform, ++++ columnar)	+	+	++	+++	+/	++	+	+	(+)	+	+
Flower size (- less than 16 mm, + more than 16 mm)	-	-	-	-	-	-	-	-	-	-	(-)
Keel shape (- rounded, + subangled, ++ angled)	-	-	-	(-)	-	+/	+/	-	++	-	-
Keel curvature (- about the middle, + below the middle, ++ lower third)	-	(-)	(-)	(-)	-	+/	(-)	-	++	-	-
Keel vestiture (upper edge) (- glabrous or ciliate, + lanate-pubescent)	-	(-)	-	-	-	(-)	+	-	-	-	-
Keel beak (- flat, + circumflexed up to 90°, ++ twisted more than 90°)	-	-	++	++	-	(-)	-	-	-	++	-
Style curvature (- rounded, + geniculate)	(-)	(-)	-	(-)	+	-	(-)	-	+	-	-
Stylar trichome distribution (+ one line, ++ two lines, +++ all around)	+	+/	++	+/	+/	++	+	+/	(+)	+++	+
Stigma (- contracted, + dilated and bifurcate)	-	-	+	-	-	-	(-)	-	-	-	-
Fruit wall (- thick, + thin)	-	+	(-)	(-)	+	-	-	-	(-)	-	-
Fruit stipe ^d (- absent/short, + long)	-	(-)	-	-	-	(-)	-	-	-	-	(-)

^a If as long, then calyx slightly bilabiate, but style rounded with a single line of hairs.

^b If keel not twisted, then keel beak blunt and wing petals as long as or longer than the keel.

^c If slightly thin-walled, then inflorescences 12 to many-flowered.

^d Short-stipitate implies that the stipe is usually up to 7 mm long; long-stipitate implies that the stipe is usually more than 7 mm long.

1. Amphitrichae clade (100% BS, PP 1.0). Ridge callosities are invariably present on the standard petal blade and claw. The keel is rounded with a twisted beak (usually twisted more than 270°). The style is invariably rounded with hairs distributed all around.
2. Grandiflorae clade (86% BS, PP 1.0). All species are either shrubs or trees. Flowers are often very large (up to 55 mm long) with prominent hypanthia. Ridge callosities are invariably present on the standard petal blade and claw, and the keel is rounded at about or below the middle with a rounded style and trichomes in one line. The fruit is invariably stipitate (often very long-stipitate; stipe up to 95 mm long in *C. laburnifolia*).
3. Glaucae clade (98% BS, PP 1.0). This group includes robust perennial herbs that become somewhat woody or rarely annuals. Ridge callosities are present on the standard petal blade and claw. The keel is rounded at about the middle, often with the upper edge crested and the beak circumflexed up to 90° or subangled below the middle with an untwisted beak. The calyx is usually more than two thirds as long as the keel or as long as the keel. The style is invariably rounded with one or two lines of hairs.
4. Incanae clade (96% BS, PP 1.0). Ridge callosities are present on the standard petal blade and claw. The calyx is deeply lobed and more or less as long as the keel with curved bracteoles ascending from the top of the pedicel (often at the base of the calyx). The keel is rounded at about the middle or angled in the lower third and the style is rounded or geniculate with one (rarely two) line of hairs. Fruits are oblong-clavate to fusiform, thin-walled (rarely thick-walled) and shortly stipitate or sessile.
5. Schizostigma clade. Three diminutive species with very small flowers and disc callosities restricted to the standard petal blade. The keel is deeply rounded at about or below the middle with a twisted beak (circumflexed 180°) and the style dilated or sometimes bifurcate at the tip with hairs in two lines.
6. Calycinae clade (100% BS, PP 1.0). Lamelliform callosities are invariably present on the standard petal blade. The calyx is usually bilabiate (rarely subequally lobed) and often as long as the keel. A subangled keel (rounded if the calyx is subequally lobed or sometimes angled in the lower third) is usually present with a twisted beak. The style is rounded or geniculate with hairs in one or two lines. The leaves are usually simple and less often compound. Bracteoles are often inserted at the base of the calyx.
7. Borealigeniculatae clade (100% BS, PP 1.0). Disc or columnar callosities are restricted to the standard petal blade. The keel is rounded or subangled below the middle with an untwisted beak and the style is invariably geniculate with one or two lines of hairs. The fruit is invariably thin-walled.
8. Crotalaria s.l. clade (Axillariflora, Crotalaria, Camp- tosepala, Dispermae and Longirostres clades, 72% BS, PP 1.0). All species have disc callosities restricted to the standard petal blade. The

keel is either rounded below the middle (rarely at about the middle) or angled in the lower third. A twisted keel beak is usually present but is absent in those species with deeply rounded keels with blunt beaks and wings that are more or less as long as or longer than the keel. The style is rounded (in those species with rounded or subangled keels) and geniculate (species with angled keels) and hairs are distributed in two lines.

The molecular phylogeny is not in exact agreement with Polhill's (1982) classification system. The following uncertainties have been resolved in this study: (1) The genus cannot be divided into two subgenera because of convergences in the position and shape of the standard petal callosities (see Le Roux, 2011). (2) The three species alliances of the *Hedriocarpae* clade (Fig. 3) are incongruent with the circumscriptions of the two subsections of section *Hedriocarpae* and no apomorphies are currently available to support a division of section *Hedriocarpae*. (3) Section *Chrysocalycinae* is paraphyletic, but three of the subsections form monophyletic groups, namely the *Stipulosae*, *Incanae* and *Glaucuae* clades. Subsection *Chrysocalycinae* is embedded within the *Hedriocarpae* clade and subsection *Tetralobocalyx* is embedded within the *Glaucuae* clade. The diversity within section *Chrysocalycinae* can only be accommodated by recircumscribing the main monophyletic groups as sections (i.e., by raising subsections to sectional status). (4) Section *Geniculatae* is paraphyletic (*Geniculatae* and *Borealigeniculatae* clades). Species with a southern African distribution (including widely distributed species) form part of the *Geniculatae* clade and the species restricted to the northern parts of northeast tropical Africa, northern Africa and the Arabian Peninsula are included in the *Borealigeniculatae* clade. (5) The *Amphitrichae* clade forms a strongly supported monophyletic group (100% BS, PP 1.00 and five synapomorphies) that does not belong in section *Crotalaria*. This clade has been described as a new section (Le Roux & Van Wyk, 2013). (6) The sectional status of section *Calycinae* is justified (Bisby & Polhill, 1973) and it should not be included in section *Crotalaria*. Concerns previously raised by Polhill (1982) with regard to the placement of *C. recta*, *C. spectabilis* and allies are here clarified—they belong to the *Calycinae* clade (represented by *C. recta* in the phylogeny). This placement is supported by the presence of lamelliform callosities on the standard petal blade. (7) Section *Crotalaria* subsection *Crotalaria* is paraphyletic (*Axillariflora*, *Camptosepala* and *Crotalaria* clades). With further study of the *Axillariflora* and *Camptosepala* clades, new apomorphies may be identified to support these clades as subsections. Furthermore, the *Camptosepala* clade is noteworthy because it includes species formerly placed in section *Hedriocarpae* but with disc callosities, stylar hairs in two lines and the absence of a twisted keel beak. Thulin (1991) was correct in suggesting that these species are misplaced in *Hedriocarpae* and that they are related to *C. emarginella* Vatke and allies of section *Crotalaria* subsection *Crotalaria*. This relationship is here confirmed by the molecular results. (8) The circumscription of Polhill's (1982) section *Dispermae* (*Dispermae* clade) received support but the section is now embedded within the *Crotalaria* s.l. clade. (9) The placement of individual species such as *C. bongensis*, *C. microcarpa*, *C. boranica*, *C. microphylla*, *C. micans*, *C. pisicarpa* and various others has also been clarified (see Le Roux, 2011).

Ansari (2008) modified Polhill's (1982) classification system to make provision for the Indian *Crotalaria* species. He used concepts from Wight & Walker-Arnott (1834), Bentham (1843) and

Baker (1876) and recognized six sections and twelve subsections (Ansari, 2006, 2008). The system focused on the Indian species and did not take into account the full morphological diversity found in Africa. Discrepancies between Polhill's (1982) and Ansari's (2006, 2008) classification systems are mainly encountered in their sections *Calycinae* and *Crotalaria* (Table 1). Ansari (2006, 2008) created four subsections within each of these sections. Molecular data supports the concept of section *Crotalaria* subsection *Longirostrae* sensu Ansari but not the other three subsections (*Bracteatae*, *Crotalaria*, *Poly-phyllae*), all of which should be included in section *Calycinae* (Table 1).

The current study provides new insights into the phylogeny of the genus *Crotalaria* and shows that several modifications to Polhill's (1982) infrageneric classification system have become necessary, not only to accommodate non-African species but also to reflect the eleven main groups or clades shown in Figure 3. These groups are quite distinct and it is unlikely that their circumscriptions will have to be modified after more detailed studies. By recognizing these as sections, new sampling strategies will become possible for the first time.

In contrast, it is not yet possible to formalise groups (e.g., subsections) within the eleven sections recognized here. Further sampling is required (to include more species, especially of sections *Hedriocarpae*, *Calycinae* and section *Crotalaria* subsection *Crotalaria*) to determine relationships within the Hedriocarpae, Calycinae, Axillariflora, Camptosepala and Crotalaria clades (possibly creating subsections).

This study revealed that specialised features used by Polhill (1968) to define his sections and subsections are not only convergent but also not always logically correlated. For example, in *C. bongensis* and *C. microcarpa* of the Hedriocarpae clade, a non-twisted keel co-occurs with a geniculate style, whereas in the Amphitrichae clade, a twisted keel co-occurs with a rounded style. It is clear that the evolution of specialised floral characters (shape and position of callosities, shape and position of the keel curvature, presence or absence of a twist in the keel, and shape and vestiture of the style) has been convergent and that the weighting of these characters has resulted in a partially artificial arrangement of species.

Taxonomic treatment

A first global infrageneric classification system is presented and formalised below, based on molecular and morphological evidence (Le Roux, 2011; Le Roux & al., 2011; Le Roux & Van Wyk, 2012). Eleven sections are recognized (derived from the clades in Fig. 3; broadened circumscriptions indicated by "s.l."): (1) section *Hedriocarpae* s.l. (Hedriocarpae clade); (2) section *Incanae* (Incanae clade, previously a subsection of section *Chrysocalycinae*); (3) section *Schizostigma* (Schizostigma clade); (4) section *Calycinae* s.l. (Calycinae clade); (5) section *Borealigeniculatae* (Borealigeniculatae clade, a new section); (6) section *Crotalaria* s.l. (including the Axillariflora, Crotalaria, Dispermae, Camptosepala and Longirostres clades); (7) section *Stipulosae* (Stipulosae clade, previously a subsection of section *Chrysocalycinae*); (8) section *Glaucae* s.l. (Glaucae clade, previously a subsection of section *Chrysocalycinae*); (9) section *Geniculatae* (Geniculatae clade); (10) section *Amphitrichae* (Amphitrichae clade, a new

section); and (11) section *Grandiflorae* (*Grandiflorae* clade). Section *Dispermae* is included in section *Crotalaria* and subsections *Tetralobocalyx* and *Chrysocalycinae* (both formerly subsections of section *Chrysocalycinae*) are included in sections *Glaucæ* and *Hedriocarpæ*, respectively. The details of the broadened circumscriptions of sections *Hedriocarpæ*, *Calycinae*, *Crotalaria* and *Glaucæ* are not indicated here but can be seen in the synonymy given below.

Key to the sections of *Crotalaria*

1. Standard petal callosities of the ridge type (Fig. 1A1); present on blade and claw 2
1. Standard petal callosities of the columnar (Fig. 1A2), disc (Fig. 1A3) or lamelliform (Fig. 1A4) types; present on blade only 7
2. Keel rounded at about the middle (Fig. 1B1, B3, B4); keel beak twisted (Fig. 1B2, B6); stylar trichomes all around distal part of style (Fig. 1F3, F6) **C. sect. *Amphitrichae***
2. Keel rounded at about or below the middle; keel beak not twisted (circumflexed up to 90°; Fig. 1B3); stylar trichomes in one or two lines (Fig. 1F1, F2, F4, F5) 3
3. Flowers usually more than 16 mm long (if shorter than 16 mm, then stipules absent); large shrubs or trees; receptacle prominent, 2–9 mm long; calyx campanulate (Fig. 1C2), never truncate; fruit oblong-clavate, often long-stipitate **C. sect. *Grandiflorae***
3. Flowers less than 16 mm long (if longer than 16 mm, then stipules present, but if stipules absent, then keel lanate along the upper edge or bracteoles inserted at the base of the calyx); mostly annuals or perennials, rarely shrubs; receptacle not prominent, less than 2 mm long (rarely up to ± 4 mm long); calyx sometimes truncate (Fig. 1C1); fruit shape variable, rarely long-stipitate 4
3. Keel with upper edge glabrous or ciliolate (Fig. 1B1–B3); keel beak sometimes circumflexed up to 90° 5
4. Keel with upper edge lanate (Fig. 1B4) to slightly tomentose (if glabrous, then stipules conspicuously cordate-falcate and asymmetrical); keel beak never twisted 6
4. Bracteoles usually inserted at the base of the calyx (Fig. 1E2); stylar trichomes invariably in one line along the upper edge of the style (Fig. 1F1, F4); calyx often truncate (Fig. 1C1) and up to half as long as the keel; petals commonly distinctly veined brown (Fig. 1C1), purple or red; keel rounded without a crest or circumflexed beak (Fig. 1B1) or rarely angled (if angled, then style geniculate; Fig. 1F8) **C. sect. *Hedriocarpæ***
5. Bracteoles variously inserted on the pedicel (rarely at the base of the calyx; Fig. 1E1); stylar trichomes in one line along the upper edge of the style (Fig. 1F1, F4) or in two lines along the upper and lower edges of the style (Fig. 1F2, F5); calyx never truncate and usually more than half as long as the keel; petals not distinctly veined purple or red (Fig. 1C2); keel rounded with a crest and sometimes circumflexed beak (Fig. 1B3) or sub-angled (Fig. 1B2) to angled in the lower half (but never with a geniculate style) **C. sect. *Glaucæ***

6. Stipules linear; bracteoles almost always curved and ascending from the pedicel or base of the calyx, filiform to ovate, persistent; calyx \pm as long as keel; fruit oblong-clavate or fusiform and thin-walled **C. sect. *Incanae***

6 Stipules expanded and leaf-like (linear or absent if leaves unifoliolate); bracteoles not curved and ascending from the pedicel, filiform to linear-lanceolate, caducous; calyx shorter than the keel; fruit cylindrical or oblong-ellipsoid and thick-walled **C. sect. *Stipulosae***

7 Keel beak not twisted (Fig. 1B5); style geniculate (Fig. 1F8); fruit subglobose or ellipsoid to oblong-obovoid; callosities of the columnar (Fig. 1A2) or rarely disc (Fig. 1A3) type 8

7. Keel beak usually twisted (Fig. 1B2 and B6; if not twisted, then calyx lobes \pm as long as the keel with wings as long or longer than the keel) (Fig. 1B6); style rounded or geniculate (Fig. 1F7 and F8 respectively); fruit shape various; callosities of the lamelliform (Fig. 1A4) or disc (Fig. 1A3) type 9

8. Keel rounded below the middle or subangled in lower third; fruit subglobose-ellipsoid to oblong-obovoid and thin-walled; distributed in northeast Tropical Africa, northern Africa and extending into the Arabian Peninsula **C. sect. *Borealigeniculatae***

9. Keel angled in the lower third; fruit oblong-ellipsoid and thick-walled (if thin-walled, then located in southern Africa); widely distributed in Africa, concentrated in southern and South Tropical Africa **C. sect. *Geniculatae***

10. Callosities lamelliform (Fig. 1A4); keel rounded (Fig. 1B1) or subangled (Fig. 1B2) below the middle, rarely angled in the lower third (Fig. 1B5, B6) and invariably with a twisted beak (Fig. 1B2, B6); calyx more than half as long as the keel to longer than the keel and deeply lobed, often bilabiate (Fig. 1D2); leaves usually simple, rarely trifoliolate **C. sect. *Calycinae***

9. Callosities disciform (Fig. 1A3); keel subangled below the middle (Fig. 1B2) or angled in the lower third (Fig. 1B5, B6) and usually with a twisted beak (Fig. 1B2, B6; if not twisted, then calyx lobes \pm as long as the keel with wings as long or longer than the keel); calyx usually shorter than the keel (if equal or longer, then keel not twisted or style dilated and bifurcate at the tip) and equally or sub-equally lobed (Fig. 1D1); leaves uni- to trifoliolate, rarely simple 10

10. Calyx as long as the keel; keel beak twisted; style dilated and bifurcate at the tip **C. sect. *Schizostigma***

10. Calyx shorter than the keel (calyx as long as or longer than keel if beak not twisted); keel beak twisted; style not dilated and bifurcate at the tip **C. sect. *Crotalaria***

1. ***Crotalaria* sect. *Hedriocarpae*** Wight & Arn., Prodr. Fl.

Ind. Orient. 1: 194. 1834, **emend. nov.** M.M. le Roux & B.-E. van Wyk – Type: *C. clavata* Wight & Arn.

= *Chrysocalyx* Guill. & Perr. in Guillemin & al., Fl. Seneg. Tent.: 157, t. 43. 1831 & 1832 = *Crotalaria* subg. *Chryso-calyx* (Guill. & Perr.) Meisn., Pl. Vasc. Gen.: Tab. Diagn. 82, Commentarius 58. 1837 – Type (designated by Cowan, 1959): *Chrysocalyx ebenoides* Guill. & Perr.

- = *Priotropis* Wight & Arn., Prodr. Fl. Ind. Orient. 1: 180. 1834 ≡ *Crotalaria* subg. *Priotropis* (Wight & Arn.) Meisn., Pl. Vasc. Gen.: Tab. Diagn. 82, Commentarius 58. 1837 ≡ *C.* subsect. *Priotropis* (Benth. ex Wight & Arn.) Polhill in Kew Bull. 22: 247. 1968 – Type: *P. cytisoides* (Roxb.) Wight & Arn. (≡ *C. cytisoides* Roxb.).
- = *Crotalaria* [unranked] *Chrysocalycinae* Benth. in London J. Bot. 2: 571. 1843; Baker in Oliver, Fl. Trop. Afr. 2: 8. 1871, pro parte ≡ *C.* ser. *Chrysocalycinae* (Benth.) Taub. in Engler & Prantl, Nat. Pflanzenfam. 3(3): 229. 1893 ≡ *C.* sect. *Chrysocalycinae* (Benth.) Baker f. in J. Linn. Soc., Bot. 42: 305. 1914 ≡ *C.* subsect. *Chrysocalycinae* (Benth.) Bisby & Polhill in New Phytol. 72: 732, 741. 1973 – Type (designated by Polhill, 1968): *C. ebenoides* (Guill. & Perr.) Walp.
- = *Crotalaria* [unranked] *Farctae* Benth. in London J. Bot. 2: 583. 1843 ≡ *C.* sect. *Farctae* (Benth.) Baker f. in J. Linn. Soc., Bot. 42: 308. 1914 – Type: *C. lupinoides* Hochst. ex Benth.
- = *Crotalaria* [unranked] *Cytisoides* Benth. in London J. Bot. 2: 591. 1843 – Type: *C. cytisoides* Hils. & Bojer ex Benth. 1843, nom. illeg., non Roxb. 1825 nec Wight 1831 (= *C. ibi-tyensis* R. Vig. & Humbert).
- = *Maria-antonia* Parl. in Maria Antonia: 3. 1844 – Type: *M. orientalis* Parl. (= *C. microcarpa* Hochst. ex Benth.).
- = *Crotalaria* sect. *Coursia* Peltier in J. Agric. Trop. Bot. Appl. 6: 267. 1959 – Type: *C. coursii* Peltier.
- = *Crotalaria* [unranked] *Macrostachyae* Benth. in London J. Bot. 2: 583. 1843 ≡ *C.* ser. *Macrostachyae* (Benth.) Taub. in Engler & Prantl, Nat. Pflanzenfam. 3(3): 227. 1893 ≡ *C.* sect. *Macrostachyae* (Benth.) Polhill in Kew Bull. 22: 261. 1968 ≡ *C.* subsect. *Macrostachyae* (Benth.) Bisby & Polhill in New Phytol. 72: 741. 1973 – Type (designated by Polhill, 1968): *C. pallida* Aiton (= *C. striata* DC.).

The two subsections of *C.* sect. *Hedriocarpae* are amalgamated and the circumscription is broadened to include *C.* sub- sect. *Chrysocalycinae*.

2. ***Crotalaria*** sect. ***Incanae*** (Benth.) Polhill in Kew Bull. 22: 217. 1968 ≡ *C.* [unranked] *Incanae* Benth. in London J. Bot. 2: 587. 1843 ≡ *C.* ser. *Incanae* (Benth.) Taub. in Engler & Prantl, Nat. Pflanzenfam. 3(3): 229. 1893 ≡ *C.* subsect. *Incanae* (Benth.) Bisby & Polhill in New Phytol. 72: 741. 1973 – Type: *C. incana* L.
- = *Phyllocalyx* A. Rich., Tent. Fl. Abyss. 1: 160. 1847 – Type: *P. quartinianus* A. Rich. (≡ *C. quartiana* A. Rich.).

The delimitation of the section follows the original sectional concept of Polhill (1968).

3. ***Crotalaria*** sect. ***Schizostigma*** Polhill in Kew Bull. 22: 297. 1968 – Type: *C. leptocarpa* Balf. f.

The circumscription of this small section (Polhill, 1968) remains the same but excludes *C. ballyi*, the only species with asymmetrical (falcate-cordate) stipules, which now forms part of *C.* sect. *Stipulosae*.

4. ***Crotalaria*** sect. ***Calycinae*** Wight & Arn. in Prodr. Fl. Ind. Orient. 1: 181. 1834, **emend. nov.** M.M. le Roux & B.-E. van Wyk – Type: *C. calycina* Schrank.
- = *Crotalaria* sect. *Alatae* Wight & Arn. in Prodr. Fl. Ind. Orient. 1: 181. 1834 ≡ *C. ser. Alatae* (Wight & Arn.) Taub. in Engler & Prantl, Nat. Pflanzenfam. 3(3): 228. 1893 ≡ *C. subsect. Alatae* (Wight & Arn.) A.A. Ansari in J. Econ. Taxon. Bot. 30: 572. 2006 – Type (designated by Polhill, 1968): *C. scabrella* Wight & Arn.
- = *Crotalaria* [unranked] *Arenariae* Benth. in London J. Bot. 2: 473. 1843 ≡ *C. sect. Arenariae* (Benth.) Baker in Hooker, Fl. Brit. India 2: 65. 1876 ≡ *C. ser. Arenariae* (Benth.) Taub. in Engler & Prantl, Nat. Pflanzenfam. 3(3): 228. 1893 – Type: *C. arenaria* Benth.
- = *Crotalaria* sect. *Bracteatae* Wight & Arn. in Prodr. Fl. Ind. Orient. 1: 183. 1834 ≡ *C. subsect. Bracteatae* (Wight & Arn.) A.A. Ansari in J. Econ. Taxon. Bot. 30: 576. 2006 Type (designated by Ansari, 2006): *C. paniculata* Willd.
- = *Crotalaria* sect. *Diffusae* Wight & Arn. in Prodr. Fl. Ind. Orient. 1: 188. 1834 ≡ *C. ser. Diffusae* (Wight & Arn.) Taub. in Engler & Prantl, Nat. Pflanzenfam. 3(3): 228. 1893 ≡ *C. subsect. Diffusae* (Wight & Arn.) A.A. Ansari in J. Econ. Taxon. Bot. 30: 573. 2006 – Type (designated by Polhill, 1968): *C. bifaria* L.
- = *Crotalaria* sect. *Erectae* Wight & Arn. in Prodr. Fl. Ind. Orient. 1: 186. 1834 ≡ *C. subsect. Retusae* Wight & Arn. in Prodr. Fl. Ind. Orient. 1: 186. 1834 – Type: *C. retusa* L.
- = *Crotalaria* sect. *Eriocarpae* Wight & Arn. in Prodr. Fl. Ind. Orient. 1: 185. 1834 ≡ *C. ser. Eriocarpae* (Wight & Arn.) Taub. in Engler & Prantl, Nat. Pflanzenfam. 3(3): 228. 1893 – Type (designated by Polhill, 1968): *C. juncea* L.
- = *Crotalaria* sect. *Fulvae* Wight & Arn. in Prodr. Fl. Ind. Orient. 1: 182. 1834 – Type: *C. fulva* Roxb.
- = *Crotalaria* sect. *Microcarpae* Wight & Arn. in Prodr. Fl. Ind. Orient. 1: 191. 1834 – Type (designated by Ansari, 2006): *C. nana* Burm. f.
- = *Crotalaria* sect. *Podocarpae* Wight & Arn. in Prodr. Fl. Ind. Orient. 1: 193. 1834 – Type (designated by Polhill, 1968): *C. orixensis* Rottler ex Willd.
- = *Crotalaria* sect. *Polyphyllae* Wight & Arn. in Prodr. Fl. Ind. Orient. 1: 194. 1834 ≡ *C. subsect. Polyphyllae* (Wight & Arn.) A.A. Ansari in J. Econ. Taxon. Bot. 30: 578. 2006 Type (designated by Polhill, 1968): *C. quinquefolia* L.
- = *Crotalaria* sect. *Sphaerocarpae* Wight & Arn. in Prodr. Fl. Ind. Orient. 1: 190. 1834 – Type (designated by Polhill, 1968): *C. biflora* L.
- = *Crotalaria* subg. *Iocaulon* Raf., New Fl. 2: 53. 1837 – **Type (designated here):** *C. sagittalis* L.
- = *Crotalaria* [unranked] *Multifoliolatae* Baker in Hooker, Fl. Brit. India 2: 84. 1876 – **Type (designated here):** *C. quinquefolia* L.
- = *Crotalaria* subsect. *Heylandiae* A.A. Ansari in J. Econ. Taxon. Bot. 30: 574. 2006 – Type: *C. hebecarpa* (DC.) Rudd.

The circumscription of this section is here broadened to include *C. spectabilis* and allies of *C. subsect. Crotalaria* (Polhill, 1982), as well as *C. subsect. Bracteatae*, *C. subsect.*

Crotalaria (sensu A.A. Ansari) and *C.* subsect. *Polyphyllae* (Ansari, 2006, 2008). *Crotalaria* subsect. *Crotalaria* as circumscribed by Ansari (2006, 2008) fits with *C.* sect. *Calycinae* but does not include the type of the genus (*C. lotifolia* L.). It is consequently excluded from the synonymy. Callosities are invariably lamelliform and therefore a useful diagnostic character.

5. ***Crotalaria*** sect. ***Borealigeniculatae*** M.M. le Roux & B.-E. van Wyk, **sect. nov.** – Type: *C. allophylla* Thulin. Sectio Borealigeniculatae sectioni Genuculatis similis est, sed carina rotundata ad subangularem curvatura circum vel infra media, lobis calycis 2–3 plo longioribus tubo et fructibus parietibus gracilibus differt (in sectione Genuculatis carina angulata curvatura in triente inferior, lobis calycis brevioribus ad longiores tubo et fructibus crassiparietibus).

A small group (*C. allophylla*, *C. boranica*, *C. microphylla*) that is morphologically similar to *C.* sect. *Genuculatae* in having untwisted keel beaks and genuculate styles. However, *C.* sect. *Borealigeniculatae* is restricted to northern areas of Africa (Egypt, Eritrea, Ethiopia, north-east Kenya, Somalia, Sudan; *C. microphylla* has a wider distribution in west and north-east tropical Africa) and the Arabian Peninsula (Saudi Arabia, Yemen). These species have rounded to subangled keel petals with the curvature at about or below the middle, columnar disc callosities, calyx lobes that are two to three times longer than the tube and subglobose-ellipsoid fruits with thin walls. Species from *C.* sect. *Genuculatae* are mostly found in southern Africa and Angola, with a few widespread. They all have angled keel petals with the curvature in the lower third, disc or columnar callosities; calyx lobes that are shorter to longer than the tube (if more than twice as long as the tube, fruit not thin-walled and species found in southern Africa) and oblong-obovoid to cylindrical fruit (less commonly subglobose) with thick valves.

6. ***Crotalaria*** L. sect. ***Crotalaria***, **emend.** M.M. le Roux & B.-E. van Wyk – Type: *C. lotifolia* L.

= *Crotalaria* sect. *Dispermae* Wight & Arn. in Prodr. Fl. Ind. Orient. 1: 191. 1834, **syn. nov.**

≡ *C.* [unranked] *Medicagineae* Benth. in London J. Bot. 2: 577. 1843 – Type: *C. medicaginea* Lam. (typified for *C.* sect. *Dispermae* by Polhill, 1968).

= *Crotalaria* [unranked] *Vulgares* Benth. in London J. Bot. 2: 592. 1843 – Type: *C. macrocarpa* E. Mey.

= *Crotalaria* [unranked] *Longirostres* Benth. in London J. Bot. 2: 580. 1843 ≡ *C.* ser. *Longirostres* (Benth.) Taub. in Engler & Prantl, Nat. Pflanzenfam. 3(3): 229. 1893 ≡ *C.* subsect. *Longirostres* (Benth.) Polhill in Kew Bull. 22: 312. 1968 - Type (designated by Polhill, 1968): *C. distans* Benth.

= *Crotalaria* [unranked] *Oocarpae* Baker in Oliver, Fl. Trop. Afr. 2: 9. 1871 – Type (designated by Polhill, 1938): *C. senegalensis* (Pers.) Bacle ex DC.

= *Crotalaria* [unranked] *Trifoliolatae* Baker in Hooker, Fl. Brit.

India 2: 81. 1879 – **Type (designated here):** *C. trifoliolum* Willd.

= *Crotalaria* [unranked] *Dispermae* Baker in Hooker, Fl. Brit. India 2: 81. 1876 – **Type (designated here):** *C. medicaginea* Lam.

- = *Crotalaria* sect. *Erectae* Wight & Arn., Prodr. Fl. Ind. Orient. 1: 186. 1834 ≡ *C.* ser. *Erectae* (Wight & Arn.) Taub. in Engler & Prantl, Nat. Pflanzenfam. 3(3): 228. 1893 – **Type (designated here):** *C. formosa* Wight & Arn.
- = *Crotalaria* sect. *Dispermae* Wight & Arn. in Prodr. Fl. Ind. Orient. 1: 191. 1834 ≡ *C.* ser. *Dispermae* (Wight & Arn.) Taub. in Engler & Prantl, Nat. Pflanzenfam. 3(3): 229. 1893 – **Type (designated here):** *C. rigida* B. Heyne ex Roth.
- = *Crotalaria* [unranked] *Fruticosae* Benth. in London J. Bot. 2: 591. 1843 ≡ *C.* ser. *Fruticosae* (Benth.) Taub. in Engler & Prantl, Nat. Pflanzenfam. 3(3): 229. 1893 – **Type (designated here):** *C. lotifolia* L.
- = *Crotalaria* sect. *Spinosa* Baker f. in J. Linn. Soc., Bot. 42: 310. 1914 – Type: *C. spinosa* Hochst. ex Benth.

Crotalaria sect. *Crotalaria* is here broadened to include *C.* sect. *Dispermae* and some species previously included in *C.* sect. *Hedriocarpae* (Axillariflora, Camptosepala, *Crotalaria*, *Dispermae* and *Longirostres* clades, Fig. 3).

7. ***Crotalaria* sect. *Stipulosae*** (Baker f.) M.M. le Roux & B.-E. van Wyk, **stat. nov.** ≡ *C.* [unranked] *Stipulaceae* Benth. in London J. Bot. 2: 588. 1843 ≡ *C.* ser. *Stipulaceae* (Benth.) Taub. in Engler & Prantl, Nat. Pflanzenfam. 3(3): 229. 1893, **syn. nov.** ≡ *C.* subsect. *Stipulosae* Baker f. in J. Linn. Soc., Bot. 42: 403. 1914 – **Type** (designated by Polhill, 1968) for *C.* subsect. *Stipulosae* Baker f.; **designated here** for *C.* [unranked] *Stipulaceae* Benth.: *C. gorensis* Guill. & Perr.

Crotalaria subsect. *Stipulosae* is raised to sectional level and the circumscription remains the same as described by Polhill (1982).

8. ***Crotalaria* sect. *Glaucæ*** (Benth.) M.M. le Roux & B.-E. van Wyk, **stat. nov.** ≡ *C.* [unranked] *Glaucæ* Benth. in London J. Bot. 2: 479. 1843 ≡ *C.* ser. *Glaucæ* (Benth.) Taub. in Engler & Prantl, Nat. Pflanzenfam. 3(3): 228. 1893, pro parte ≡ *C.* subsect. *Glaucæ* (Benth.) Polhill in Kew Bull. 22: 230. 1968 – Type: *C. glauca* Willd.
- = *Crotalaria* sect. *Tetralobocalyx* Harms in Bot. Jahrb. Syst. 54: 383. 1917 ≡ *C.* subsect. *Tetralobocalyx* (Harms) Bisby & Polhill in New Phytol. 72: 741. 1973, **syn. nov.** Type: *C. nigricans* Baker.

The circumscription of this section is slightly broadened to include not only *C.* subsect. *Glaucæ* (Benth.) Polhill but also the monotypic *C.* subsect. *Tetralobocalyx* (Harms) Bisby & Polhill (the latter characterised by a four-lobed calyx). In the molecular phylogeny, *C. nigricans* is shown to be embedded within *C.* sect. *Glaucæ*.

9. ***Crotalaria* sect. *Geniculatae*** Polhill in Kew Bull. 22: 294. 1968 ≡ *C.* [unranked] *Sphaerocarpae* Benth. in London J. Bot. 2: 579. 1843, non *C.* sect. *Sphaerocarpae* Wight & Arn. 1834 ≡ *C.* sect. *Sphaerocarpae* (Benth.) Baker f. in J. Linn. Soc. Bot. 42: 273. 1914, nom. illeg., non Wight & Arn. 1834 – Type: *C. sphaerocarpa* Perr. ex DC.

The circumscription of this section (Polhill 1968, 1982) is slightly narrowed to exclude species with ridge callosities (e.g., *C. bongensis* and *C. microcarpa*) that form part of *C. sect. Hedriocarpae* and three species with thin-walled fruits that are restricted to North-East Tropical Africa and Arabia, now included in *C. sect. Borealigeniculatae*.

10. ***Crotalaria*** sect. ***Amphitrichae*** M.M. le Roux & B.-E. van Wyk in Syst. Bot. 38: 645. 2013 – Type: *C. colorata* Schinz
= *Crotalaria* [unranked] *Oliganthae* Benth. in London J. Bot. 2: 573. 1843 ≡ *C. subsect. Oliganthae* (Benth.) Baker f. in J. Linn. Soc., Bot. 42: 386. 1914 – **Type (designated here):** *C. humilis* Eckl. & Zeyh.

A group of seven species (*C. colorata*, *C. excisa*, *C. humilis*, *C. meyerana*, *C. pearsonii* and two new species) with a unique combination of floral characters, of which the amphistylar hairs is the most useful (trichomes entirely surround the apical part of the style). This character is found elsewhere only in *C. monophylla* (*C. sect. Geniculatae*). The new section is the only one with a restricted distribution, limited to the west coast of South Africa and southern Namibia.

11. ***Crotalaria*** sect. ***Grandiflorae*** (Baker f.) Polhill in Kew Bull. 22: 201. 1968 ≡ *C. subsect. Grandiflorae* Baker f. in J. Linn. Soc., Bot. 42: 314. 1914 – Type: *Crotalaria laburnifolia* L.
= *Clavulium* Desv. in Ann. Sci. Nat. (Paris) 9: 407. 1826 ≡ *Crotalaria* subg. *Clavulium* (Desv.) Meisn., Pl. Vasc. Gen.: Tab. Diagn. 82, Commentarius 58. 1837 – Type: *Clavulium pedunculosum* (Desv.) Desv. (≡ *Crotalaria pedunculosa* Desv.).

This section remains the same as circumscribed by Polhill (1968, 1982).

Acknowledgements

The following people and institutions are gratefully acknowledged: the National Research Foundation and the University of Johannesburg for financial support; the staff of CEN, ICN, JRAU, K, L, MEL, MO, NBG, P, SI, UPS and WIND for organizing herbarium loans and DNA samples; Dr. G. Koorsen (University of Johannesburg) for translating the diagnosis into Latin; Dr. J. Iganci (Federal University of Rio Grande do Sul) for obtaining literature, Dr. A. Magee (SANBI) for help with molecular analyses; the Barcode of Life Data Systems and Guelph University for DNA sequencing (of the barcoding gene regions, *matK* and *rbcLa*); staff of the Jodrell Laboratory at Kew for DNA extractions and the staff of the Molecular and Systematics Laboratory at the University of Johannesburg for the use of facilities and technical support.

Literature cited

- Ansari, A.A.** 2006. Taxonomic studies on the genus *Crotalaria* L. in India – II: Infrageneric classification. *J. Econ. Taxon. Bot.* 30: 570–582.
- Ansari, A.A.** 2008. *Crotalaria L. in India*. Dehra-Dunn: Bishen Singh Mahendra Pal Singh.
- Bain, J.F. & Jansen, R.K.** 2006. A chloroplast DNA hairpin structure provides useful phylogenetic data within tribe Senecioneae (Asteraceae). *Canad. J. Bot.* 84: 862–868. <http://dx.doi.org/10.1139/b06-057>
- Baker, E.G.** 1914. The African species of *Crotalaria*. *J. Linn. Soc., Bot.* 42: 241–425.
- Baker, J.G.** 1876. *Crotalaria*. Pp. 65–85 in: Hooker, J.D. (ed.), *Flora of British India*, vol. 2. London: Reeve & Co.
- Bentham, G.** 1843. Enumeration of the Leguminosae indigenous to southern Asia and central and southern Africa XV. *Crotalaria*. *London J. Bot.* 2: 472–481, 559–593.
- Bisby, F.A.** 1970. The evaluation and selection of characters in angio-sperm taxonomy: An example from *Crotalaria*. *New Phytol.* 69: 1149–1160. <http://dx.doi.org/10.1111/j.1469-8137.1970.tb02495.x>
- Bisby, F.A.** 1973. The role of taxometrics in angiosperm taxonomy I. Empirical comparisons of methods using *Crotalaria* L. *New Phytol.* 72: 699–726. <http://dx.doi.org/10.1111/j.1469-8137.1973.tb04420.x>
- Bisby, F.A. & Polhill, R.M.** 1973. The role of taxometrics in angio-sperm taxonomy II. Parallel taximetric and orthodox studies in *Crotalaria* L. *New Phytol.* 72: 727–742. <http://dx.doi.org/10.1111/j.1469-8137.1973.tb04421.x>
- Boatwright, J.S., Le Roux, M.M., Wink, M., Morozova, T. & Van Wyk, B.-E.** 2008a. Phylogenetic relationships of tribe Crotalarieae (Fabaceae) inferred from DNA sequences and morphology. *Syst. Bot.* 33: 752–761. <http://dx.doi.org/10.1600/036364408786500271>
- Boatwright, J.S., Savolainen, V., Van Wyk, B.-E., Schutte-Vlok, A.L., Forest, F. & Van der Bank, M.** 2008b. Systematic position of the anomalous genus *Cadia* and the phylogeny of the tribe Podalyrieae (Fabaceae). *Syst. Bot.* 33: 133–147. <http://dx.doi.org/10.1600/036364408783887500>
- Boatwright, J.S., Wink, M. & Van Wyk, B.-E.** 2011. The generic concept of *Lotononis* (Crotalarieae, Fabaceae): Reinstatement of the genera *Euchlora*, *Leobordea* and *Listia* and the new genus *Ezoloba*. *Taxon* 60: 161–177.
- Cowan, R.S.** 1959. Leguminosae of the western hemisphere. *Taxon* 8: 58–60. <http://dx.doi.org/10.2307/1216031>
- Downie, S.R. & Katz-Downie, D.S.** 1996. A molecular phylogeny of Apiaceae subfamily Apioideae: Evidence from nuclear ribosomal DNA internal transcribed spacer sequences. *Amer. J. Bot.* 83: 234–251. <http://dx.doi.org/10.2307/2445943>
- Doyle, J.J. & Doyle, J.L.** 1987. A rapid DNA isolation procedure for small quantities of fresh leaf tissue. *Phytochem. Bull.* 19: 11–15.
- Edwards, D., Horn, A., Taylor, D., Savolainen, V. & Hawkins, J.A.** 2008. DNA barcoding of a large genus, *Aspalathus* L. (Fabaceae). *Taxon* 57: 1317–1327.

- Farris, J.S., Källersjö, M., Kluge, A.G. & Bult, C.** 1995. Testing significance of incongruence. *Cladistics* 10: 315–319. <http://dx.doi.org/10.1111/j.1096-0031.1994.tb00181.x>
- Felsenstein, J.** 1985. Confidence limits on phylogenies: An approach using the bootstrap. *Evolution* 39: 783–791. <http://dx.doi.org/10.2307/2408678>
- Fitch, W.M.** 1971. Towards defining the course of evolution: Minimum change for a specified tree topology. *Syst. Zool.* 20: 406–416. <http://dx.doi.org/10.2307/2412116>
- Flores, A.S., Corrêa, A.M., Forni-Martins, E.R. & Tozzi, A.M.G.A.** 2006. Chromosome numbers in Brazilian species of *Crotalaria* (Leguminosae, Papilionoideae) and their taxonomic significance. *Bot. J. Linn. Soc.* 151: 271–277. <http://dx.doi.org/10.1111/j.1095-8339.2006.00479.x>
- Harvey, W.H.** 1862. Leguminosae. Pp. 39–47 in: Harvey, W.H. & Sonder, O.W. (eds.), *Flora capensis*, vol. 2. Dublin: Hodges, Smith & Co.
- Hepper, F.N.** 1958. Papilionaceae, *Crotalaria*. Pp. 544–553 in: Keay, R.W.J. (ed.), *Flora of west tropical Africa*. London: Crown Agents.
- Huelsenbeck, J.P. & Ronquist, F.** 2001. MRBAYES: Bayesian inference of phylogeny. *Bioinformatics* 17: 754–755. <http://dx.doi.org/10.1093/bioinformatics/17.8.754>
- Jianqiang, L., Sun, H., Polhill, R.M. & Gilbert, M.G.** 2010. Crotalarieae: *Crotalaria*. Pp. 105–117 in: Wu, Z.Y., Raven, P.H. & Hong, D.Y. (eds.), *Flora of China*, vol. 10. Beijing: Science Press; St. Louis: Missouri Botanical Garden Press.
- Kress, W.J. & Erickson, D.L.** 2007. A two-locus global DNA barcode for land plants: The coding *rbcl* gene complements the non-coding *psbA-trnH* spacer region. *PLoS One* 2: e508. <http://dx.doi.org/10.1371/journal.pone.0000508>
- Kress, W.J., Erickson, D.L., Jones, F.A., Swenson, N.G., Perez, R., Sanjur, O. & Bermingham, E.** 2009. Plant DNA barcodes and a community phylogeny of a tropical forest dynamics plot in Panama. *Proc. Natl. Acad. Sci. U.S.A.* 106: 18621–18626. <http://dx.doi.org/10.1073/pnas.0909820106>
- Lahaye, R., Van der Bank, M., Bogarin, D., Warner, J., Pupulin, F., Gigot, G., Maurin, O., Duthoit, S., Barraclough, T.G. & Savolainen, V.** 2008. DNA barcoding the floras of biodiversity hotspots. *Proc. Natl. Acad. Sci. U.S.A.* 105: 2923–2928. <http://dx.doi.org/10.1073/pnas.0709936105>
- Le Roux, M.M.** 2011. *A taxonomic study of the genus Crotalaria L. (Fabaceae, tribe Crotalarieae) and a modified infrageneric classification system*. Ph.D. thesis, University of Johannesburg, South Africa.
- Le Roux, M.M. & Van Wyk, B.-E.** 2012. The systematic value of flower structure in *Crotalaria* and related genera of the tribe Crotalarieae (Fabaceae). *Flora* 207: 414–426. <http://dx.doi.org/10.1016/j.flora.2012.02.005>
- Le Roux, M.M. & Van Wyk, B.-E.** 2013. A taxonomic revision of *Amphitrichae*, a new section of *Crotalaria* (Fabaceae). *Syst. Bot.* 38: 638–652. <http://dx.doi.org/10.1600/036364413X670296>
- Le Roux, M.M., Van Wyk, B.-E., Boatwright, J.S. & Tilney, P.M.** 2011. The systematic significance of morphological and anatomical variation in fruits of *Crotalaria* and related genera of tribe Crotalarieae (Fabaceae). *Bot. J. Linn. Soc.* 165: 84–106. <http://dx.doi.org/10.1111/j.1095-8339.2010.01098.x>

- Milne-Redhead, E.** 1961. Miscellaneous notes on African species of *Crotalaria* L. *Kew Bull.* 15: 157–167. <http://dx.doi.org/10.2307/4115786>
- Pandey, A., Singh, R., Sharma, S.K. & Bhandari, D.C.** 2010. Diversity assessment of useful *Crotalaria* species in India for plant genetic resources management. *Genet. Resources Crop Evol.* 57: 461–470. <http://dx.doi.org/10.1007/s10722-009-9517-0>
- Polhill, R.M.** 1968. Miscellaneous notes on African species of *Crotalaria* L. II. *Kew Bull.* 22: 169–348. <http://dx.doi.org/10.2307/4107767>
- Polhill, R.M.** 1976. Genisteeae (Adans.) Benth. and related tribes (Leguminosae). *Bot. Syst.* 1: 143–368.
- Polhill, R.M.** 1982. *Crotalaria in Africa and Madagascar*. Rotterdam: A.A. Balkema.
- Posada, D. & Crandall, K.A.** 1998. Modeltest: Testing the model of DNA substitution. *Bioinformatics* 13: 817–818. <http://dx.doi.org/10.1093/bioinformatics/14.9.817>
- Rambaut & Drummond.** 2009. Tracer. <http://tree.bio.ed.ac.uk/download.html?name=tracer&version=v1.5&id=80&num=2>
- Ratnasingham, S. & Hebert, P.D.N.** 2007. BOLD: The Barcode of Life Data System (www.barcodinglife.org). *Molec. Ecol. Notes* 7: 355–364. <http://dx.doi.org/10.1111/j.1471-8286.2007.01678.x>
- Reeves, G., Chase, M.W., Goldblatt, P., Fay, M.F., Cox, A.V., Lejeune, B. & Souza-Chies, T.** 2001. Molecular systematics of Iridaceae: Evidence from four plastid DNA regions. *Amer. J. Bot.* 88: 2074–2087. <http://dx.doi.org/10.2307/3558433>
- Ronquist, F. & Huelsenbeck, J.P.** 2003. MRBAYES 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics* 19: 1572–1574. <http://dx.doi.org/10.1093/bioinformatics/btg180>
- Sang, T., Crawford, D.J. & Stuessy, T.F.** 1997. Chloroplast DNA phylogeny, reticulate evolution, and biogeography of *Paeonia* (Paeoniaceae). *Amer. J. Bot.* 84: 1120–1136. <http://dx.doi.org/10.2307/2446155>
- Schreiber, A.** 1970. Fabaceae. Pp. 16–28 in: Merxmüller, H. (ed.), *Prodromus einer Flora von Südwestafrika*, Lfg. 60. Lehre: J. Cramer.
- Seelanan, T., Schnabel, A. & Wendel, J.F.** 1997. Congruence and consensus in the cotton tribe (Malvaceae). *Syst. Bot.* 22: 259–290. <http://dx.doi.org/10.2307/2419457>
- Sun, Y., Skinner, D.Z., Liang, G.H. & Hulbert, S.H.** 1994. Phylogenetic analysis of sorghum and related taxa using internal transcribed spacers of nuclear ribosomal DNA. *Theor. Appl. Genet.* 89: 26–32. <http://dx.doi.org/10.1007/BF00226978>
- Swofford, D.L.** 2002. PAUP*: Phylogenetic analysis using parsimony (*and other methods). Sunderland, Massachusetts: Sinauer.
- Tamura, K., Dudley, J., Nei, M. & Kumar, S.** 2007. MEGA4: Molecular evolutionary genetics analysis (MEGA) software version 4.0. *Molec. Biol. Evol.* 24: 1596–1599. <http://dx.doi.org/10.1093/molbev/msm092>
- Thulin M.** 1991. Two new species of *Crotalaria* (Leguminosae) from Somalia. *Nordic J. Bot.* 11: 97–101. <http://dx.doi.org/10.1111/j.1756-1051.1991.tb01804.x>
- Torre, A.R.** 1962. *Crotalaria*. Pp. 6–76 in: Exell, A.W. & Fernandes, A. (eds.), *Conspectus Florae Angolensis*, vol. 3. Lisboa: Junta de Investigações do Ultramar.
- Van Wyk, B.-E.** 1991. A review of the tribe Crotalarieae (Fabaceae). *Contr. Bolus Herb.* 13: 265–288. [http://dx.doi.org/10.1016/S0305-1978\(03\)00083-8](http://dx.doi.org/10.1016/S0305-1978(03)00083-8)

- Van Wyk, B.-E.** 2003. The value of chemosystematics in clarifying relationships in the genistoid tribes of papilionoid legumes. *Bio-chem. Syst. Ecol.* 31: 875–884.
- Van Wyk, B.-E.** 2005. Tribe Crotalariaeae. Pp. 273–281 in: Lewis, G., Schrire, B., Mackinder, B. & Lock, M. (eds.), *Legumes of the World*. Kew: Royal Botanic Gardens.
- Van Wyk, B.-E. & Schutte, A.L.** 1989. Taxonomic relationships amongst some genera of Leguminosae tribe Crotalariaeae and *Argyrolobium* (Genisteae). *Kew Bull.* 44: 397–423. <http://dx.doi.org/10.2307/4110360>
- Van Wyk, B.-E. & Schutte, A.L.** 1995. Phylogenetic relationships in the tribes Podalyrieae, Liparieae and Crotalariaeae. Pp. 283–308 in: Crisp, M. & Doyle, J. (eds.), *Advances in legume systematics*, vol. 7. Kew: Royal Botanic Gardens.
- Van Wyk, B.-E. & Verdoorn, G.** 1990. Alkaloids as taxonomic characters in the tribe Crotalariaeae (Fabaceae). *Biochem. Syst. Ecol.* 18: 503–515. [http://dx.doi.org/10.1016/0305-1978\(90\)90122-V](http://dx.doi.org/10.1016/0305-1978(90)90122-V)
- Van Wyk, B.-E., Venter, M. & Boatwright, J.S.** 2010. A revision of the genus *Bolusia* (Fabaceae, Crotalariaeae). *S. African J. Bot.* 76: 86–94. <http://dx.doi.org/10.1016/j.sajb.2009.08.010>
- Verdoorn, I.C.** 1928. A revision of the Crotalarias of South and South- East Tropical Africa. *Bothalia* 2: 371–420.
- Werle, E., Schneider, C., Renner, M., Volker, M. & Fiehnet, W.** 1994. Convenient single-step, one tube purification of PCR products for direct sequencing. *Nucl. Acids Res.* 22: 4354–4355. <http://dx.doi.org/10.1093/nar/22.20.4354>
- White, T.J., Bruns, T., Lee, S. & Taylor, J.** 1990. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. Pp. 315–322 in: Innis, M.A., Gelfand, M.A., Sninsky, J.J. & White, T.J. (eds.), *PCR protocols*. New York: Academic Press.
- Whitlock, B.A., Hale, A.M. & Groff, P.A.** 2010. Intraspecific inversions pose a challenge for the *trnH-psbA* plant DNA barcode. *PLoS ONE* 5: e11533. <http://dx.doi.org/10.1371/journal.pone.0011533>
- Wight, R. & Walker-Arnott, G.A.** 1834. *Prodromus florae peninsulae Indiae orientalis*. London: Parburry, Allen & Co.
- Wilczek, R.** 1953a. Papilionaceae Genisteae Congolanae Novae (*Robynsiophyton*, *Crotalaria*, *Argyrolobium*). *Bull. Jard. Bot. État Bruxelles* 23: 125–211.
- Wilczek, R.** 1953b. *Crotalaria*. Pp. 43–273 in: Robyns, W., Staner, P., Demaret, F., Germain, R., Gilbert, G., Hauman, L., Homès, M., Jurion, F., Lebrun, J., Van den Abeele, M. & Boutique, R. (eds.), *Flore du Congo Belge*, vol. 4. Brussels: Institut National pour l'Étude Agronomique du Congo belge.
- Yoder, A.D., Irwin, J.A. & Payseur, B.A.** 2001. Failure of the ILD to determine data combinability for slow loris phylogeny. *Syst. Biol.* 50: 408–424. <http://dx.doi.org/10.1080/106351501300318003>